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**ARTICLE** in EXPERT OPINION ON THERAPEUTIC TARGETS · JULY 2013

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# **EXPERT OPINION**

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The role of mitochondrial dysfunctions due to oxidative and nitrosative stress in the chronic pain or chronic fatigue syndromes and fibromyalgia patients: peripheral and central mechanisms as therapeutic targets?

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Introduction: Chronic fatigue syndrome (CFS) and fibromyalgia (FM) are characterized by persistent pain and fatigue. It is hypothesized that reactive oxygen species (ROS), caused by oxidative and nitrosative stress, by inhibiting mitochondrial function can be involved in muscle pain and central sensitization as typically seen in these patients.

Areas covered: The current evidence regarding oxidative and nitrosative stress and mitochondrial dysfunction in CFS and FM is presented in relation to chronic widespread pain. Mitochondrial dysfunction has been shown in leukocytes of CFS patients and in muscle cells of FM patients, which could explain the muscle pain. Additionally, if mitochondrial dysfunction is also present in central neural cells, this could result in lowered ATP pools in neural cells, leading to generalized hypersensitivity and chronic widespread pain. Expert opinion: Increased ROS in CFS and FM, resulting in impaired mitochondrial function and reduced ATP in muscle and neural cells, might lead to chronic widespread pain in these patients. Therefore, targeting increased ROS by antioxidants and targeting the mitochondrial biogenesis could offer a solution for the chronic pain in these patients. The role of exercise therapy in restoring mitochondrial dysfunction remains to be explored, and provides important avenues for future research in this area.

Keywords: ATP, central sensitization, chronic pain, mitochondria, muscle, nitric oxide, NMDA receptor, peroxynitrite, spinal cord, superoxide

Expert Opin. Ther. Targets [Early Online]

#### 1. Introduction

## 1.1 Defining chronic fatigue syndrome and fibromyalgia

Chronic fatigue syndrome (CFS) is a debilitating and complex disorder, characterized by extreme fatigue [1]. The population prevalence of CFS is between 0.2 and 2.6% (with > 75% female patients [2]) and little is known about the etiology of the illness, making prevention and treatment challenging. In addition to the chronic fatigue, widespread and persistent pain is common in individuals with CFS [3-6].



## Article highlights.

- There is evidence for increased oxidative and nitrosative
- Mitochondria are both the source and the target of increased oxidative and nitrosative stress.
- Mitochondrial dysfunctions are shown in muscle cells of
- Mitochondrial dysfunctions in muscle cells may cause muscle pain, due to increased muscle damage and decreased aerobic capacity.
- If mitochondrial dysfunctions are also present in the cells of the CNS, this may lead to increased sensitivity and thus central sensitization.
- Possible therapeutic targets encompass increasing the antioxidant level or optimizing mitochondrial function.

This box summarizes key points contained in the article

The syndrome is largely overlapping with fibromyalgia (FM), a chronic pain syndrome that has been defined by widespread pain for > 3 months and the presence of additional symptoms like disturbed sleep, emotional distress, and pronounced fatigue [7,8].

Patients with FM and the majority of patients with CFS suffer from myalgias and chronic widespread pain. Evidence is growing for central sensitization as the underlying mechanism explaining chronic pain in CFS and FM [9]. But the mechanism behind the onset of central sensitization seems to be multifactorial. It is hypothesized that reactive oxygen species (ROS), caused by oxidative and nitrosative stress, by inhibiting mitochondrial function can be involved in muscle (peripheral) pain and central sensitization.

#### 1.2 Oxidative and nitrosative stress

Both syndromes are often preceded by a physical trauma, such as from an accident or from surgery, psychological stress, viral, parasitic or bacterial infections, episodes of strenuous physical activity, etc. [10,11]. Following inflammatory stimuli, the production of oxygen radicals (ROS, oxidative and nitrosative stress), H<sub>2</sub>O<sub>2</sub> (peroxides), 2O<sub>2</sub> (superoxide) and ONOO (peroxynitrite) is increased.

There is evidence for increased oxidative stress in CFS and FM, with higher oxidized low-density lipoproteins (LDL) and elevated protein carbonyl levels found in blood samples [12-19]. There is also evidence for damage caused by oxidative stress in CFS and FM. The serum IgM antibodies to fatty acids, such as phosphatidyl-inositol (Pi) and malondialdehyde (MDA), which are by-products of lipid peroxidation, are significantly greater in CFS patients compared to healthy controls. This shows that CFS is characterized by an IgM-related immune response directed against disrupted lipid membrane components and byproducts of lipid peroxidation, which are normally not detected by the immune system, but due to oxidative damage, have become immunogenic. Thus, the oxidative

damage in CFS may have functional consequences: disorders in Pi may have biological effects by interfering with intracellular signaling processes, and the damage to membrane fatty acids may render the peroxidized membrane bilayer more rigid, thus inducing changes in different membrane (including receptor) functions [20].

Inflammation is also accompanied by nitrosative stress due to overactivated iNOS (inducible nitric oxide synthase) resulting in excessive nitrogen monoxide (NO) and peroxynitrite (ONOO<sup>-</sup>) formation by activated neutrophils and monocytes. Both nitrogen monoxide and peroxynitrite levels are reported to be increased in blood peripheral lymphocytes of CFS patients [20]. Peroxynitrite reacts with and inactivates several of the enzymes in mitochondria leading to energy metabolism dysfunction [21,22]. A central tenet of the peroxynitrite hypothesis is that once levels are elevated, there are several potential positive feedback mechanisms which may act to amplify and sustain these elevated levels. The most important thing here is that the mitochondria are both the source and the target of peroxynitrite, sustaining a vicious circle [23].

There is evidence in other different myopathies and peripheral neuropathies [24] that mitochondria contribute to the pathogenesis and/or pathophysiology. In CFS and FM, the role of mitochondria in the chronic widespread pain of these patients remains obscure and underreported. Nevertheless, we hypothesize that mitochondrial function in muscle, blood and spinal cord is impaired in CFS and FM due to increased oxidative and nitrosative stress and that these impairments may lead, among others, to peripheral sources of nociception and central sensitization, explaining the chronic widespread pain and other systemic complaints in these patients. The present report will provide an overview of the existing evidence regarding mitochondrial dysfunction in these patients and explain how these abnormalities may lead to the chronic pain seen in these patients. Furthermore, possible therapeutic targets will be discussed.

## 2. Current evidence

# 2.1 Mitochondrial dysfunctions in white blood cells and skeletal muscle of CFS patients?

The available CFS data concerning mitochondrial dysfunction were obtained mainly in two targeted tissues: white blood cells and muscle cells. In white blood cells data are more sound than in the muscle tissue. In white blood cells several researchers reported decreased mitochondrial function resulting in less ATP [25-27]. However evaluating these results more in depth, some authors discussed that the main cause is decreased citrate synthase content, which is an expression of the number of mitochondria [26]. Others find a decreased activity of the different enzymes (complexes) expressed relatively to the citrate synthase content. This means that the enzymes are less active additionally to the decreased number of mitochondria [25,27].



A limited number of studies addressing muscular mitochondrial function in CFS patients are available, but data are conflicting. Some did not found any difference in mitochondrial enzyme activity in CFS patients compared to a matched control group, but only a decreased mitochondrial content [28]. Other studies have shown abnormal mitochondrial degeneration [27,29,30] and severe deletions of oxidative genes in mitochondrial DNA (mtDNA) [27]. These changes result in reduced ATP synthesis and increased lactate concentrations [29]. The most important difference between these studies are different inclusion criteria resulting in a larger heterogeneity of the studied population and the used methodology, for instance, by measuring plasma creatine kinase levels, which is an indirect method [25].

In conclusion, real progress in establishing the etiopathogenesis of CFS as a ROS-dependent process with clear impact on mitochondrial function will only be possible with definitive evidence that an excess of free radicals in CFS muscles (measured in muscle biopsies) is directly influencing the mitochondrial function. At the moment this is fundamentally lacking in the current literature.

# 2.2 Mitochondrial dysfunction in white blood cells and skeletal muscle of FM patients?

In FM patients the literature is more sound concerning mitochondrial dysfunction. Several authors report mitochondrial dysfunction in white blood cells as well as in muscle biopsies [31,32]. In these patients abnormalities in white blood and muscle cell mitochondria were detected with electron microscopy, namely mitochondria with irregular cristae, single fiber defects of cytochrome-c-oxidase, and deletions of the mitochondrial genome [33].

The degenerative muscle changes in membranes, mitochondria, and capillary vessels may be related to defects in membrane ion channeling, oxygen and metabolite transport, and ATP production via oxidative phosphorylation. The decrease in activity of the oxidative enzymes, 3-hydroxy CoA dehydrogenase, citrate synthase, and cytochrome oxidase, support the proposal of defects in oxidative metabolism and ATP synthesis [34].

# 2.3 Muscular mitochondrial dysfunctions leading to widespread peripheral muscle pain

Muscular mitochondrial dysfunctions are known to cause (exercise-induced) peripheral muscle pain, weakness, fatigability, exercise intolerance, acidosis, etc. [24]. If mitochondria become dysfunctional when they are stressed by one or more stimuli, cells can undergo apoptosis. Mitochondria contain cytochrome c and apoptogens, which enter the cytoplasm and activate caspase, which in turn causes cellular damage and induce nuclear fragmentation.

In FM patients data supporting this view are scarce. In order to show apoptosis and abnormal cell death in FM, Sprott et al. investigated DNA fragmentation in the muscle

tissue and ultra-structural analysis by electron microscopic features of the muscle in FM patients. DNA fragmentation has been detected to be significantly higher in patients with FM compared to the nuclei in healthy controls [35]. In some genetic connective tissue disorders like Bethlem and Ullrich disease, which are associated with severe hypermobility and have a complaint pattern (pain and fatigue) which is largely overlapping with FM [36], mitochondrial dysfunction and increased apoptosis in skeletal muscle have been reported [37,38].

Furthermore, it has been proposed that ROS damage can increase permeability of transition pores in the mitochondrial inner membrane, leading to a simultaneous collapse of mitochondrial membrane potential and the elimination of dysfunctional mitochondria by mitophagy, as evidenced by the findings of mitophagy in blood mononuclear cells of FM patients. "Mitophagy" is described as the selective removal of mitochondria by autophagy during development and under pathologic conditions. And while normal autophagy is functional, extensive mitophagy comprises cell functionality and may contribute to the pathophysiology of

Finally, it seems that mitochondrial dysfunction is related to the circulating cytokine levels in patients with FM and thus contributes to the pathophysiology of FM. Inversely, reducing oxidative stress in patients with FM improved mitochondrial function, reduced cytokines levels, and reduced pain scores and tender points [39]. It is indeed shown that lipid peroxidation in blood cells (due to mitochondrial ROS) is significantly correlated to pain scores, fatigue, stiffness, etc., in patients with FM [40].

But besides increased inflammation and mitophagy in FM, other mechanisms leading to peripheral pain due to possible mitochondrial dysfunction in CFS or FM are not yet clear. To date, it is not known whether mitochondrial dysfunction could, for example, lead to muscle damage and reduced aerobic capacity in CFS or FM, leading to muscle pain and peripheral sensitization (by, e.g., acidosis, hypoxia and byproducts of muscle damage) and the typical pain exacerbation after exercise.

# 2.4 Mitochondrial dysfunctions in central nervous system leading to central sensitization and chronic pain

In FM and CFS, a body of evidence is available for central sensitization as the underlying mechanism of chronic pain [41-45]. This heightened general pain sensitivity may be sustained and amplified by continuous upgrading of peripheral nociceptive input [33]. The proposed basis for this enhancement of secondary hyperalgesia is the activation of the N-methyl-D-aspartate (NMDA) receptors. The NMDA receptors in the dorsal horn of the spinal cord and periaqueductal gray have been demonstrated to play a role in the development of central sensitization [46].

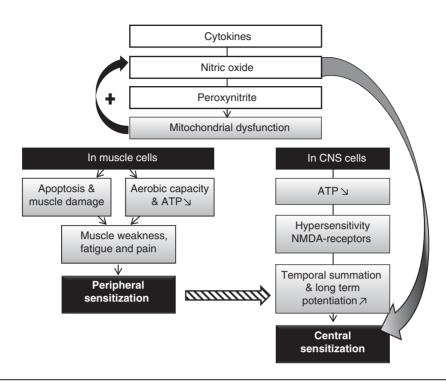


Figure 1. Relation of oxidative and nitrosative stress together with mitochondrial dysfunction to central sensitization.

It is well known that when neurons containing NMDA (N-methyl-D-aspartic acid) receptors have lowered ATP pools their NMDA receptors become hypersensitive to stimulation due to lowering of the plasma membrane potential, leading to an immediate increase in NMDA activity and stimulate NMDA-dependent temporal summation of pain longterm potentiation, thus producing longer term increases in synaptic activity. Due to the interaction with peroxynitrite ATP production, this possibly explains the contribution to central sensitization [47].

Earlier studies implicated increased sensitivity of central NMDA receptors as playing a primary role in FM, as evidenced by a significant reduction in symptoms among a large subset of patients in response to low doses of ketamine, a noncompetitive NMDA receptor antagonist [48]. Furthermore, ketamine attenuates pain, pain thresholds, referred pain and temporal summation when administered intravenously to patients with FM [49,50].

Another emerging concept in pain research is that oxidative stress play a critical role in chronic pain due to central sensitization. Elevated spinal ROS levels by increased production of mitochondrial superoxide lead to central sensitization and consequently pain without peripheral nerve injury or tissue inflammation [51]. Superoxide can cause peripheral and central sensitization and alter nociception [52], resulting in hyperalgesia mediated by both local and spinal oxidant mechanisms. Also NO has, for example, the capability of inducing peripheral and central sensitization by reducing receptor thresholds [53] and is able to reduce the inhibitory activity of

the central nervous system leading to central sensitization of dorsal horn neurons [54]. Evidence for the involvement of ROS in pain has been provided in a number of studies mainly by demonstrating the antinociceptive effect of some antioxidants [55,56], superoxide dismutase (SOD; neutralizes superoxide) mimetics [57] and inhibitors of electron transport chain complexes [58]. Furthermore, increased levels of ROS or their by-products are shown in specific tissues in animal models of persistent pain [59] and SOD inhibitors further enhanced secondary capsaicin-induced hyperalgesia [57]. Pain modulation can be accomplished by altering superoxide in the spinal cord [57]. Joseph and Levine demonstrated a role of the mitochondrial electron transport chain in neuropathic and some forms of inflammatory pain. The contribution of the mitochondrial electron transport chain in neuropathic pain is ATP-dependent [58].

However, this hypothesis with an interaction of oxidative and nitrosative stress, mitochondrial function and chronic pain due to central sensitization has not yet been studied in CFS.

Figure 1 presents a possible mitochondrial-related mechanism underlying central sensitization in CFS and FM.

# 3. Conclusion

To conclude, it seems plausible that oxidative and nitrosative stress together with mitochondrial dysfunction play an important role in both syndromes. Nevertheless, the exact mechanisms and the clinical relevance require stronger evidence.



Table 1. Therapeutic targets for oxidative stress and mitochondrial dysfunction in CFS and FM.

Objective	Possible tools
Antioxidant level /	Coenzyme 10
	Carnosine
Mitochondrial biogenesis /	Resveratrol
Mitochondrial content/enzymes /	Exercise (cave NO!)

∠: Increase; NO: Nitric oxide.

Especially, the relation of oxidative and nitrosative stress together with mitochondrial dysfunction to central sensitization has not been examined yet. Given the consistent and overwhelming evidence supporting a cardinal role for central sensitization in both FM and CFS, this is an important shortcoming of our current understanding of these syndromes. Further research should focus on these lacunas, because the present rationale offers interesting therapeutic targets in these populations: targeting the antioxidant status and optimizing mitochondrial function.

## 4. Expert opinion

As mentioned above, the presence of mitochondrial dysfunction in CFS as well as FM patients is a relevant plausible hypothesis, based on the current evidence, which potentially explains pain and fatigue in these patients. Mitochondrial dysfunction in CFS and FM patients in muscle, peripheral as well as central nervous tissue, can be induced by a large amount of ROS. These oxygen radicals are produced by the mitochondria themselves. The increased amount of free radicals in this tissue is a consequence of a negative balance between ROS-production of the mitochondria, on the one hand, and the antioxidant content in the tissue, on the other hand. The relative rates of ROS production and decomposition determine their steady state concentration and ultimately, their potential to cause tissue injury.

It is possible that the production of ROS is increased due to an increased frequency of action potentials registered in the neurons of the pain system (peripheral as well as central) due to a traumatic injury or infection which often precedes the syndromes. Due to the injury or infection, different cells will become necrotic and several humoral factors, such as bradykinine and potassium, will leak into the environment, possibly targeting nociceptors of the pain neurons. In this case there is a constant imbalance at the level of the membrane potential which has to be restored by Na/K-ATPase pumps. These pumps are ATP-dependent, which stresses the mitochondrial function, resulting in an increased production of oxygen radicals. This hypothesis is supported by Mattson and Liu who reported that mitochondria in axons and presynaptic terminals provide sources of ATP to drive the ion pumps that are concentrated in these structures, for rapid restoration of ion gradients following depolarization and neurotransmitter release [60].

Mitochondria may also play important roles in the regulation of synaptic function because of their ability to regulate calcium levels and ROS production. ROS generated in response to synaptic activity are known to contribute to the regulation of long-term structural and functional changes in neurons. The high-energy demands of synapses, together with their high levels of ROS production, place them at risk during conditions of increased stress, which occur in aging, neurodegenerative disorders such as Alzheimer's and Parkinson's diseases, and after acute traumatic and ischemic insults.

Besides an increased ROS production, it is also possible that the levels of several antioxidants in the tissue are lowered [61]. The elimination of strong oxidants is a nonenzymatic process and is done by vitamin C, vitamin E, ubiquinone, carnosine, etc. The elimination of less reactive species is done by specific enzymes, present in the cytosol, peroxisomes or mitochondria. The mitochondrion has its own set of enzymes and other antioxidants. The mitochondrial matrix contains Mn-superoxide dismutase (SOD-2) and the intermembrane space contains the cytosolic isoform of superoxide dismutase (Cu, Zn-SOD or SOD-1) [62,63].

Besides increased ROS production and lowered levels of antioxidants, inflammatory substances, such as TNF-α and interleukins, can also have a direct impact on the mitochondrial function in the inflamed environment. In liver tissue, Weidinger et al. reported that inflammation, due to hypoxia, induced mitochondrial dysfunction [64]. In CFS and FM patients, increased biomarkers of inflammation have been reported [65], and it can therefore be hypothesized that increased levels of inflammation may lead to mitochondrial

Based on the aforementioned evidence and knowledge, different therapeutic targets can be defined, as presented in Table 1.

To counter the cause of the mitochondrial dysfunction, therapy can focus on keeping up the levels of antioxidants and on mitochondrial antioxidant function.

In FM patients as well as in CFS patients, several authors reported decreased levels of Coenzyme Q10 and several minerals as zinc, copper, magnesium and manganese, which would be related to pain complaints [66-69]. Cordero reported that supplementation of Coenzyme Q10 had positive effects on mitochondrial function by decreasing oxidative stress and thereby having a positive impact on headache symptoms, tender points, VAS score and the Fibromyalgia Impact Questionnaire in FM patients [16,39,70]. In juvenile FM patients, supplementation also reduced self-reported fatigue [13]. No studies evaluating the effect of Coenzyme Q10 supplementation on pain in CFS are available to the best of our knowledge. Further clinical trials focusing on the antioxidant status in these patients and focusing on supplementation with antioxidants with interest for oxidative stress, mitochondrial function measured in blood cells and muscle tissue and evaluating association with several complaints, mainly pain and fatigue, are required.

An interesting antioxidant is carnosine ( $\beta$ -alanyl-L-histidine), a dipeptide of the amino acids β-alanine and histidine. It is highly concentrated in muscle and central nervous

Carnosine has been proven to scavenge reactive oxygen species (ROS) as well as  $\alpha$ - $\beta$  unsaturated aldehydes formed from peroxidation of cell membrane fatty acids during oxidative stress.

In animal models carnosine has been shown to retard cancer growth and protect against alcohol-induced oxidative stress as well as ethanol-induced chronic liver damage. Carnosine is also neuroprotective against permanent cerebral ischemia in mice [71].

Carnosine has been shown to counteract peroxynitritedependent protein alterations such as tyrosine nitration and thereby cell damage [72]. Boldyrev et al. [71] reported in two recent clinical trials that carnosine levels were depressed in Parkinson's disease and after brain stroke, and that supplementation resulted in a decrease in blood plasma protein carbonyls and lipid hydroperoxides (ROS indicators) and this was accompanied with increased cerebral blood flow, locomotor and cognitive function. It should be interesting to evaluate the content of carnosine in muscle tissue and blood in CFS and FM patients and if carnosine is decreased supplementation studies should be done.

A third element in the possible solutions of decreasing ROS and increasing antioxidant capacity in the cell are medicationspecific targeting the mitochondrial biogenesis. As an example we name resveratrol or esveratrol (3,5,4'-trihydroxy-trans-stilbene) which is a stilbenoid, a type of natural phenol, and a phytoalexin produced naturally by several plants, especially the roots of the Japanese Knotweed, from which it is extracted commercially.

The effects of resveratrol are currently a topic of numerous animal and human studies. In mouse and rat experiments, anticancer, anti-inflammatory, blood sugar-lowering and other beneficial cardiovascular effects of resveratrol have been reported [73].

It could be speculated that these types of medication can increase the content of mitochondria and thereby the content of antioxidants, which would help to decrease the amount of ROS, but further research is required in patients with FM and CFS.

Finally, due to the fact that these patients have decreased mitochondrial content due to the pathology but also due to physical deconditioning, it is desirable to increase this by increasing physical activity, for example, by physical exercise In healthy individuals, aerobic, strength, combined exercise training and interval training induces morphological, metabolic and gene-regulating adaptations in skeletal muscle including increases in muscle respiratory capacity, components of the mitochondrial respiratory chain, ATP synthase, and enzymes of both the Kreb's cycle and β-oxidation pathways [74]. The effect of exercise training on antioxidant func tion of mitochondria remains however element of debate. Physical activity normally causes an increase in NO production [75,76]. If this effect would be reinforced by the already elevated amounts of NO this could explain the typical pain exacerbation, malaise and the delayed recovery after physical activity as typically seen in FM and CFS patients. The results of Jammes et al. indeed confirm a lengthened and accentuated oxidative stress after exercise in patients with CFS, sufficient to explain muscle pain and postexertional malaise [77]. Additionally, a subgroup of CFS patients presents a significantly lower ATP synthesis rate during recovery, indicating impaired mitochondrial oxidative phosphorylation [78].

Furthermore, pathological overproduction of NO will decrease oxygen consumption [79] and increase anaerobic glycolysis (lactate production) by modulating mitochondrial res piration [80] and iron metabolism [81]. These effects in addition to the oxidative damage of cell membranes [82], structural proteins as actin and DNA [83], may compromise exercise capacity and worsen physical activity responses. So, although the increase of NO may be one among more beneficial effects of exercise in healthy people, in CFS and FM patients more research is required to study the effect of exercise on NO production, to elucidate the benefits or the dangers of exercise [84]. Additionally, further investigation into the possible positive effects of exercise training on mitochondrial function has to be elaborated and certainly the most optimal form with respect to the complaints.

## **Declaration of interest**

The authors state no conflict of interest and have received no payment in preparation of this manuscript.



#### **Bibliography**

Papers of special note have been highlighted as either of interest (•) or of considerable interest ( o o ) to readers

- Fukuda K, Straus SE, Hickie I, et al. The chronic fatigue syndrome: a comprehensive approach to its definition and study. International Chronic Fatigue Syndrome Study Group. Ann Intern Med 1994;121(12):953-9
- Prins JB, van der Meer JW, Bleijenberg G. Chronic fatigue syndrome. Lancet 2006;367(9507):346-55
- Buchwald D. Fibromyalgia and chronic 3. fatigue syndrome: similarities and differences. Rheum Dis Clin North Am 1996;22(2):219-43
- 4 Goldenberg DL, Simms RW, Geiger A, Komaroff AL. High frequency of fibromyalgia in patients with chronic fatigue seen in a primary care practice. Arthritis Rheum 1990;33(3):381-7
- 5. Meeus M, Nijs J, Meirleir KD. Chronic musculoskeletal pain in patients with the chronic fatigue syndrome: a systematic review. Eur J pain (London, England) 2007;11(4):377-86
- Nijs J, Vaes P, McGregor N, et al. Psychometric properties of the Dutch Chronic Fatigue Syndrome-Activities and Participation Questionnaire (CFS-APQ). Phys Ther 2003;83(5):444-54
- Wolfe F, Smythe HA, Yunus MB, et al. The American College of Rheumatology 1990 Criteria for the Classification of Fibromyalgia. Report of the Multicenter Criteria Committee. Arthritis Rheum 1990;33(2):160-72
- Wolfe F, Clauw DJ, Fitzcharles MA, et al. The American College of Rheumatology preliminary diagnostic criteria for fibromyalgia and measurement of symptom severity. Arthritis Care Res 2010;62(5):600-10
- Nijs J, Meeus M, Van Oosterwijck J, et al. In the mind or in the brain? Scientific evidence for central sensitisation in chronic fatigue syndrome. Eur J Clin Invest 2012;42(2):203-12
- Theorell T, Blomkvist V, Lindh G, Evengard B. Critical life events, infections, and symptoms during the year preceding chronic fatigue syndrome (CFS): an examination of CFS patients and subjects with a nonspecific life crisis. Psychosom Med 1999;61(3):304-10

- Van Houdenhove B, Neerinckx E, 11. Onghena P, et al. Premorbid "overactive" lifestyle in chronic fatigue syndrome and fibromyalgia. An etiological factor or proof of good citizenship? J Psychosom Res 2001;51(4):571-6
- 12. Chung CP, Titova D, Oeser A, et al. Oxidative stress in fibromyalgia and its relationship to symptoms. Clin Rheumatol 2009;28(4):435-8
- Miyamae T, Seki M, Naga T, et al. Increased oxidative stress and coenzyme Q10 deficiency in juvenile fibromyalgia: amelioration of hypercholesterolemia and fatigue by ubiquinol-10 supplementation. Redox Rep 2013;18(1):12-19
- 14. Neyal M, Yimenicioglu F, Aydeniz A, et al. Plasma nitrite levels, total antioxidant status, total oxidant status, and oxidative stress index in patients with tension-type headache and fibromyalgia. Clin Neurol Neurosurg 2013;115(6):736-40
- Cordero MD, de Miguel M, 15 Carmona-Lopez I, et al. Oxidative stress and mitochondrial dysfunction in fibromyalgia. Neuro Endocrinol Lett 2010;31(2):169-73
- Evidence for oxidative stress and mitochondrial dysfunction in FM in relation to symptomatology.
- 16 Cordero MD, Cano-Garcia FJ, Alcocer-Gomez E, et al. Oxidative stress correlates with headache symptoms in fibromyalgia: coenzyme Q(1)(0) effect on clinical improvement. PLoS One 2012;7(4):e35677
- Kennedy G, Spence VA, McLaren M, 17. et al. Oxidative stress levels are raised in chronic fatigue syndrome and are associated with clinical symptoms. Free Radical Biol Med 2005;39(5):584-9
- Evidence for oxidative stress in CFS in relation to symptomatology.
- Cordero MD, De Miguel M, 18. Moreno Fernandez AM, et al. Mitochondrial dysfunction and mitophagy activation in blood mononuclear cells of fibromyalgia patients: implications in the pathogenesis of the disease. Arthritis Res Ther 2010;12(1):R17
- 19. Ozgocmen S, Ozyurt H, Sogut S, et al. Antioxidant status, lipid peroxidation and nitric oxide in fibromyalgia: etiologic

- and therapeutic concerns. Rheumatol Int 2006;26(7):598-603
- 20 Maes M, Twisk FN. Chronic fatigue syndrome: harvey and Wessely's (bio) psychosocial model versus a bio (psychosocial) model based on inflammatory and oxidative and nitrosative stress pathways. BMC Med 2010:8:35
- Beckman JS, Koppenol WH. Nitric oxide, superoxide, and peroxynitrite: the good, the bad, and ugly. Am J Physiol 1996;271(5 Pt 1):C1424-37
- 22. Radi R, Rodriguez M, Castro L, Telleri R. Inhibition of mitochondrial electron transport by peroxynitrite. Arch Biochem Biophys 1994;308(1):89-95
- Pall ML. Elevated, sustained peroxynitrite levels as the cause of chronic fatigue syndrome. Med Hypotheses 2000;54(1):115-25
- In-depth hypothesis about the role of NO in the pathophysiology of CFS.
- Nardin RA, Johns DR. Mitochondrial 24 dysfunction and neuromuscular disease. Muscle Nerve 2001;24(2):170-91
- Vermeulen RC, Kurk RM, Visser FC, et al. Patients with chronic fatigue syndrome performed worse than controls in a controlled repeated exercise study despite a normal oxidative phosphorylation capacity. J Transl Med 2010;8:93
- 26. Booth NE, Myhill S, McLaren-Howard J. Mitochondrial dysfunction and the pathophysiology of Myalgic Encephalomyelitis/Chronic Fatigue Syndrome (ME/CFS). Int J Clin Exp Med 2012;5(3):208-20
- Evidence for mitochondrial dysfunction in CFS in relation to symptomatology.
- Myhill S, Booth NE, McLaren-Howard J. Chronic fatigue syndrome and mitochondrial dysfunction. Int J Clin Exp Med 2009;2(1):1-16
- Evidence for mitochondrial dysfunction in CFS in relation to symptomatology.
- 28. Smits B, van den Heuvel L, Knoop H, et al. Mitochondrial enzymes discriminate between mitochondrial disorders and chronic fatigue syndrome. Mitochondrion 2011;11(5):735-8



- Behan WM, More IA, Behan PO. 29 Mitochondrial abnormalities in the postviral fatigue syndrome. Acta Neuropathol 1991;83(1):61-5
- 30 Vecchiet L, Montanari G, Pizzigallo E, et al. Sensory characterization of somatic parietal tissues in humans with chronic fatigue syndrome. Neurosci Lett 1996;208(2):117-20
- 31. Gerdle B, Forsgren MF, Bengtsson A, et al. Decreased muscle concentrations of ATP and PCR in the quadriceps muscle of fibromyalgia patients - A (31) P-MRS study. Eur J Pain (London, England) 2013; Epub ahead of print
- 32. Cordero MD, Moreno-Fernandez AM, Carmona-Lopez MI, et al. Mitochondrial dysfunction in skin biopsies and blood mononuclear cells from two cases of fibromyalgia patients. Clin Biochem 2010;43(13-14):1174-6
- Park IH, Niermann KI, Olsen N. 33. Evidence for metabolic abnormalities in the muscles of patients with fibromyalgia. Curr Rheumatol Rep 2000;2(2):131-40
- Pongratz DE, Spath M. Morphologic aspects of fibromyalgia. Z Rheumatol 1998;57(Suppl 2):47-51
- Sprott H, Salemi S, Gay RE, et al. Increased DNA fragmentation and ultrastructural changes in fibromyalgic muscle fibres. Ann Rheum Dis 2004;63(3):245-51
- Rombaut L, Malfait F, De Paepe A, 36. et al. Impairment and impact of pain in female patients with Ehlers-Danlos syndrome: a comparative study with fibromyalgia and rheumatoid arthritis. Arthritis Rheum 2011;63(7):1979-87
- 37. Grumati P, Coletto L, Sandri M, Bonaldo P. Autophagy induction rescues muscular dystrophy. Autophagy 2011;7(4):426-8
- Grumati P, Coletto L, Schiavinato A, et al. Physical exercise stimulates autophagy in normal skeletal muscles but is detrimental for collagen VI-deficient muscles. Autophagy 2011;7(12):1415-23
- 39. Cordero MD, Diaz-Parrado E, Carrion AM, et al. Is inflammation a mitochondrial dysfunction-dependent event in fibromyalgia? Antioxid Redox Signal 2013;18(7):800-7
- Cordero MD, Alcocer-Gomez E, 40. Cano-Garcia FJ, et al. Clinical symptoms in fibromyalgia are better associated to lipid peroxidation levels in blood

- mononuclear cells rather than in plasma. PLoS One 2011;6(10):e26915
- Meeus M, Nijs J. Central sensitization: a biopsychosocial explanation for chronic widespread pain in patients with fibromyalgia and chronic fatigue syndrome. Clin Rheumatol 2007;26(4):465-73
- Explanation about the mechanisms and the role of central sensitization in the pain of patients with CFS and FM.
- Arendt-Nielsen L, Graven-Nielsen T. Central sensitization in fibromyalgia and other musculoskeletal disorders. Curr Pain Headache Rep 2003;7(5):355-61
- Staud R, Robinson ME, Price DD. 43 Temporal summation of second pain and its maintenance are useful for characterizing widespread central sensitization of fibromyalgia patients. J Pain 2007;8(11):893-901
- Staud R, Smitherman ML. Peripheral and central sensitization in fibromyalgia: pathogenetic role. Curr Pain Headache Rep 2002;6(4):259-66
- Staud R, Vierck CJ, Cannon RL, et al. Abnormal sensitization and temporal summation of second pain (wind-up) in patients with fibromyalgia syndrome. Pain 2001;91(1-2):165-75
- Baranauskas G, Nistri A. Sensitization of pain pathways in the spinal cord: cellular mechanisms. Prog Neurobiol 1998;54(3):349-65
- Tan EC, Janssen AJ, Roestenberg P, et al. Mitochondrial dysfunction in muscle tissue of complex regional pain syndrome type I patients. Eur J Pain (London, England) 2011;15(7):708-15
- Wood PB. A reconsideration of the relevance of systemic low-dose ketamine to the pathophysiology of fibromyalgia. J Pain 2006;7(9):611-14
- Sorensen J, Bengtsson A, Backman E, et al. Pain analysis in patients with fibromyalgia. Effects of intravenous morphine, lidocaine, and ketamine. Scand J Rheumatol 1995;24(6):360-5
- Graven-Nielsen T, Aspegren Kendall S, Henriksson KG, et al. Ketamine reduces muscle pain, temporal summation, and referred pain in fibromyalgia patients. Pain 2000;85(3):483-91
- Kim HY, Chung JM, Chung K. Increased production of mitochondrial superoxide in the spinal cord induces

- pain behaviors in mice: the effect of mitochondrial electron transport complex inhibitors. Neurosci Lett 2008;447(1):87-91
- 52 Wang ZQ, Porreca F, Cuzzocrea S, et al. A newly identified role for superoxide in inflammatory pain. J Pharmacol Exp Ther 2004;309(3):869-78
- Goupille P, Jayson MI, Valat JP, Freemont AJ. The role of inflammation in disk herniation-associated radiculopathy. Semin Arthritis Rheum 1998;28(1):60-71
- Vikman KS, Hill RH, Backstrom E, et al. Interferon-gamma induces characteristics of central sensitization in spinal dorsal horn neurons in vitro. Pain 2003;106(3):241-51
- Kim HK, Park SK, Zhou JL, et al. Reactive oxygen species (ROS) play an important role in a rat model of neuropathic pain. Pain 2004;111(1-2):116-24
- Lee I, Kim HK, Kim JH, et al. The role of reactive oxygen species in capsaicin-induced mechanical hyperalgesia and in the activities of dorsal horn neurons. Pain 2007;133(1-3):9-17
- 57 Schwartz ES, Kim HY, Wang J, et al. Persistent pain is dependent on spinal mitochondrial antioxidant levels. J Neurosci 2009;29(1):159-68
- Rationale of mitochondrial involvement in central sensitization.
- Joseph EK, Levine JD. Mitochondrial electron transport in models of neuropathic and inflammatory pain. Pain 2006;121(1-2):105-14
- Schwartz ES, Lee I, Chung K, Chung JM. Oxidative stress in the spinal cord is an important contributor in capsaicin-induced mechanical secondary hyperalgesia in mice. Pain 2008;138(3):514-24
- Mattson MP, Liu D. Energetics and oxidative stress in synaptic plasticity and neurodegenerative disorders. Neuromolecular Med 2002;2(2):215-31
- Role of ROS in neuroplasticity.
- Desai KM, Chang T, Wang H, et al. Oxidative stress and aging: is methylglyoxal the hidden enemy? Can J Physiol Pharmacol 2010;88(3):273-84
- Fischer LR, Igoudjil A, Magrane J, et al. 62. SOD1 targeted to the mitochondrial intermembrane space prevents motor



- neuropathy in the Sod1 knockout mouse. Brain 2011:134(Pt 1):196-209
- Igoudjil A, Magrane J, Fischer LR, et al. In vivo pathogenic role of mutant SOD1 localized in the mitochondrial intermembrane space. I Neurosci 2011;31(44):15826-37
- Weidinger A, Dungel P, Perlinger M, et al. Experimental data suggesting that inflammation mediated rat liver mitochondrial dysfunction results from secondary hypoxia rather than from direct effects of inflammatory mediators. Frontiers Physiol 2013;4:138
- Maes M, Twisk FN, Ringel K. Inflammatory and cell-mediated immune biomarkers in myalgic encephalomyelitis/ chronic fatigue syndrome and depression: inflammatory markers are higher in myalgic encephalomyelitis/chronic fatigue syndrome than in depression. Psychother Psychosom 2012;81(5):286-95
- 66 Werbach MR. Nutritional strategies for treating chronic fatigue syndrome. Altern Med Rev 2000;5(2):93-108
- Maes M, Mihaylova I, Kubera M, et al. Coenzyme Q10 deficiency in myalgic encephalomyelitis/chronic fatigue syndrome (ME/CFS) is related to fatigue, autonomic and neurocognitive symptoms and is another risk factor explaining the early mortality in ME/CFS due to cardiovascular disorder. Neuro Endocrinol Lett 2009;30(4):470-6
- Cordero MD, Moreno-Fernandez AM, 68 deMiguel M, et al. Coenzyme Q10 distribution in blood is altered in patients with fibromyalgia. Clin Biochem 2009;42(7-8):732-5
- Kim YS, Kim KM, Lee DJ, et al. Women with fibromyalgia have lower levels of calcium, magnesium, iron and manganese in hair mineral analysis. J Korean Med Sci 2011;26(10):1253-7
- Cordero MD, Cotan D, del-Pozo-Martin Y, et al. Oral coenzyme Q10 supplementation improves clinical symptoms and recovers pathologic alterations in blood mononuclear cells in a fibromyalgia patient. Nutrition 2012;28(11-12):1200-3

- Boldyrev AA, Stvolinsky SL, 71. Fedorova TN, Suslina ZA, Carnosine as a natural antioxidant and geroprotector: from molecular mechanisms to clinical trials. Rejuvenation Res 2010;13(2-3):156-8
- 72. Derave W, Everaert I, Beeckman S, Baguet A. Muscle carnosine metabolism and beta-alanine supplementation in relation to exercise and training. Sports Med (Auckland, NZ) 2010;40(3):247-63
- Xu Q, Si LY. Resveratrol role in cardiovascular and metabolic health and potential mechanisms of action. Nutr Res 2012:32(9):648-58
- Kessler HS, Sisson SB, Short KR. The 74. potential for high-intensity interval training to reduce cardiometabolic disease risk. Sports Med (Auckland, NZ) 2012;42(6):489-509
- Brown MD, Srinivasan M, Hogikyan RV, et al. Nitric oxide biomarkers increase during exercise-induced vasodilation in the forearm. Int J Sports Med 2000;21(2):83-9
- Green DJ, Maiorana A, O'Driscoll G, Taylor R. Effect of exercise training on endothelium-derived nitric oxide function in humans. J Physiol 2004;561(Pt 1):1-25
- Jammes Y, Steinberg JG, Mambrini O, et al. Chronic fatigue syndrome: assessment of increased oxidative stress and altered muscle excitability in response to incremental exercise. J Intern Med 2005;257(3):299-310
- Lane RJ, Barrett MC, Taylor DJ, et al. Heterogeneity in chronic fatigue syndrome: evidence from magnetic resonance spectroscopy of muscle. Neuromuscul Disord 1998;8(3-4):204-9
- 79. Shen W, Hintze TH, Wolin MS. Nitric oxide. An important signaling mechanism between vascular endothelium and parenchymal cells in the regulation of oxygen consumption. Circulation 1995;92(12):3505-12
- 80. Tatsumi T, Matoba S, Kawahara A, et al. Cytokine-induced nitric oxide production inhibits mitochondrial energy production

- and impairs contractile function in rat cardiac myocytes. J Am Coll Cardiol 2000;35(5):1338-46
- Qian ZM, Xiao DS, Ke Y, Liao QK. Increased nitric oxide is one of the causes of changes of iron metabolism in strenuously exercised rats. Am J Physiol Regul Integr Comp Physiol 2001;280(3):R739-43
- 82. Fulle S, Pietrangelo T, Mancinelli R, et al. Specific correlations between muscle oxidative stress and chronic fatigue syndrome: a working hypothesis. J Muscle Res Cell Motil 2007;28(6):355-62
- Radak Z, Pucsok J, Mecseki S, et al. 83. Muscle soreness-induced reduction in force generation is accompanied by increased nitric oxide content and DNA damage in human skeletal muscle. Free Radic Boil Med 1999;26(7-8):1059-63
- Meeus M, Van Eupen I, Hondequin J, 84. et al. Nitric oxide concentrations are normal and unrelated to activity level in chronic fatigue syndrome: a case-control study. In vivo (Athens, Greece) 2010;24(6):865-9

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