

Themed Section: Epigenetics and Therapy

REVIEW Epigenetics in the perioperative period

P Lirk¹, H Fiegl², N C Weber¹ and M W Hollmann¹

¹Department of Anaesthesiology, Academic Medical Center, University of Amsterdam, Amsterdam, The Netherlands, and ²Department of Gynaecology and Obstetrics, Innsbruck Medical University, Innsbruck, Austria

Correspondence

Philipp Lirk, Department of Anaesthesiology, Academic Medical Center, University of Amsterdam, Amsterdam, The Netherlands. E-mail: p.lirk@amc.uva.nl

Received

1 May 2014 **Revised** 13 July 2014 **Accepted** 18 July 2014

The perioperative period is characterized by profound changes in the body's homoeostatic processes. This review seeks to address whether epigenetic mechanisms may influence an individual's reaction to surgery and anaesthesia. Evidence from animal and human studies suggests that epigenetic mechanisms can explain many facets of susceptibility to acute and chronic pain, making them potential therapeutic targets. Modern pain management is still based upon opiates, and both the developmental expression of opioid receptors and opioid-induced hyperalgesia have been linked to epigenetic mechanisms. In general, opiates seem to increase global DNA methylation levels. This is in contrast to local anaesthetics, which have been ascribed a global demethylating effect. Even though no direct investigations have been carried out, the potential influence of epigenetics on the inflammatory response that follows surgery seems a promising area for research. There is a considerable body of evidence that supports the involvement of epigenetics in the complex process of wound healing. Epigenetics is an important emerging research topic in perioperative medicine, with a huge potential to positively influence patient outcome.

LINKED ARTICLES

This article is part of a themed section on Epigenetics and Therapy. To view the other articles in this section visit http://dx.doi.org/10.1111/bph.2015.172.issue-11

Abbreviations

GAD, glutamic acid decarboxylase; MeCP2, methyl-CpG-binding protein 2; OIH, opiate-induced hyperalgesia; SNP, single-nucleotide polymorphism

Tables of Links

TA	ARGETS	LIGANDS
μ	opioid receptor	Bupivacaine
D	NA methyltransferase	Buprenorphine
Hi	stone acetyltransferase	Chemokine CCL7
Hi	stone deacetylases	GABA
Hi	stone deacetylase 7	JQ1
TR	PA1 ion channel	Ketamine
		Lidocaine
		Remifentanil

These Tables list key protein targets and ligands in this article which are hyperlinked to corresponding entries in http:// www.guidetopharmacology.org, the common portal for data from the IUPHAR/BPS Guide to PHARMACOLOGY (Pawson *et al.*, 2014) and are permanently archived in the Concise Guide to PHARMACOLOGY 2013/14 (Alexander *et al.*, 2013a,b,c).

Epigenetics is concerned with the stability and expression of information contained in human DNA. The three principal mechanisms that govern the transcription of genes are the modification of histone structure, methylation of DNA regions and nucleosome positioning (Portela and Esteller, 2010). In human disease, the epigenetic signature has been





Figure 1

Summary of the main epigenetic mechanisms relevant for the perioperative period with green and blue colours designating experimental and human studies respectively. Chemokine CCL7; ; TRPA1 ion channel; prolif, proliferation.

proposed both as a biomarker (Teschendorff *et al.*, 2010) and as a therapeutic target (Navada *et al.*, 2014). Epigenetics help explain why events early in life exert profound influences on how the body will react to a similar stimulus years or even decades later (Martin *et al.*, 2010). Also, epigenetic traits are heritable, meaning that alterations in an epigenetic signature and the corresponding phenotype can sometimes be discerned generations later. One of the prototype scenarios is that of the Dutch Famine Winter of 1944–1945, a severe exogenous stressor that led to periconceptional epigenetic changes that are still detectable now, 60 years later (Heijmans *et al.*, 2008). Therefore a strong, specific or persistent enough stressor has the potential to change epigenetic markers and influence body function for years, decades and potentially in coming generations.

The perioperative period in modern medicine is unique. Surgical and anaesthetic techniques, however refined they may be, still impose considerable stress on the human body. But patient monitoring is more comprehensive and provides more safeguards than ever before, while there are countless, continuous interventions to improve outcome. Antibiotics, pain killers, hypnotics, muscle relaxants, anti-emetics, haemostatic drugs, vasopressors, inotropes and local anaesthetics are examples of drugs given preoperatively, often simultaneously. Drugs are not only given in the operating room; many of these drugs are administered for days after surgery. Given the profound physiological changes elicited by surgery and anaesthesia, we will explore whether surgery or anaesthesia may influence epigenetic signatures and so modify short- and long-term outcomes, and whether epigenetic risk factors may, at least in part, determine an individual's reaction to surgery (Bain and Shaw, 2012; Naguib et al., 2012).

This review focuses on some research fields within epigenetics that relate to the perioperative period, including susceptibility to acute and chronic pain, the effects of opiates and local anaesthetics, and potential epigenetic roles in the stress response to surgery, wound healing, and cancer recurrence (Figure 1).

Pain

Despite many advances in pain research, outcome after surgery is still bad for patients in many acute and chronic categories (Kehlet et al., 2006; Correll et al., 2014). Especially after prototype surgeries such as amputation and thoracotomy, between 50 and 80% of patients suffer from chronic pain. The prevalence of phantom pain following surgical amputation is high. A recent study confined to patients undergoing lower limb amputation for peripheral vascular disease reported phantom limb pain in 79% of patients (Richardson et al., 2006). Other survey data describe figures of around 75% (Kern et al., 2009). For amputations incurred during wartime, reports from the U.S. Department of Veteran Affairs cite similar numbers (72–76%) (Reiber et al., 2010). The reason why some patients develop ongoing pain and in others the pain resolves is still unknown. However, part of this risk is very likely to be genetic. Acute pain thresholds, in contrast to chronic pain levels, are less genetically determined (with estimated heritability scores of 22-55%) (Norbury et al., 2007). In contrast, mono- versus di-zygotic twins studies show a heritable component to the risk of developing persistent pain of up to 60% (Nyman et al., 2011). Such data clearly suggest a genetic predisposition for those who develop chronic pain following a precipitating incident, such as limb amputation and attendant nerve damage, and some preliminary insights have been gained. For example, the incidence of phantom pain post-limb amputation in

Israeli war veterans ranges from 56 to 78%, depending on the genotype of the patient at one single-nucleotide polymorphism (SNP) in *KCNS1* (rs734784, P = 0.0001). In addition the haplotype of *KCNS1* identified by this SNP was also associated with pain outcome in five out of six human populations tested (overall *P* value of association with higher pain risk = 1.14 E-08) (Costigan *et al.*, 2010).

However, in addition to genetic factors, which may determine baseline risk, epigenetic factors are increasingly seen as important in the transition from acute to chronic pain (Buchheit et al., 2012). This field of research was initiated by animal studies demonstrating an association of chronic pain with the epigenetic modification of specific genes. Zhang and colleagues investigated epigenetic contributions to the development of chronic pain in complete Freund's adjuvant injection and spinal nerve ligation. Here it was shown that, after induction of persistent inflammatory and neuropathic pain, the expression of GAD65 was epigenetically suppressed by histone hypoacetylation, decreasing inhibition in the GABA synaptic pathway. Conversely, inhibition of histone deacetylase increased GAD65 activity, led to normalized GABA transmission, and ultimately resulted in less sensitivity to pain. This effect was absent in mice knocked out for the gene encoding GAD65 (Zhang et al., 2011). In a model of partial sciatic nerve ligation, Imai and colleagues demonstrated IL-6dependent demethylation of the H3 histone at Lys²⁷, resulting in an increased expression of the chemokine CCL7 (MCP3) in glial cells. They hypothesized that this interaction would promote neuron-glial interactions, determining chronic neuropathic pain (Imai et al., 2013).

A study by Tajerian and colleagues probably has the most far-reaching implications. They demonstrated altered DNA methylation in the prefrontal cortex, in a mouse model of peripheral nerve injury associated with persistent neuropathic pain (Tajerian *et al.*, 2013). This study showed that the consequences of peripheral nerve injury reach far into the CNS. Notably, animals that received environmental enrichment therapy to alleviate pain showed a resolution of epigenetic alterations in parallel with decreases in pain behaviour. Another facet of epigenetic involvement in pain syndromes is the fact that epigenetic traits are heritable. In the case of stress-induced visceral hypersensitivity, the epigenetic methylation signature associated with this experimental irritable bowel disease could still be detected in offspring two generations later (van den Wijngaard *et al.*, 2013).

In humans, increased DNA methylation levels in peripheral blood leukocytes have been linked to the severity of chronic pain in patients on long-term treatment using potent opioids (Doehring *et al.*, 2013). A recent investigation in monozygotic twins found that the degree of heat pain sensitivity correlated with the methylation status of the gene that encodes the TRPA-1 ion channel (Bell *et al.*, 2014). This ion channel transforms thermal stimuli into heat pain (Julius, 2013). To explain the complex susceptibility and resilience to developing chronic pain, the concept of pain vulnerability has been proposed by Denk and co-authors, linking genetic and epigenetic risk factors (Denk *et al.*, 2014).

In conclusion, epigenetic mechanisms seem to explain many facets of acute and chronic pain susceptibility. However, further work needs to be done to explain the detailed role of perioperative factors. Opiates are a fundamental component of perioperative analgesic regimens, and one of the oldest classes of drugs in existence. They can be subdivided into drugs with differential affinity for one of four major receptor families, that is μ , κ , δ or nociceptin receptors (Chiou *et al.*, 2004; Pasternak, 2004), all belonging to the family of GPCRs.

The expression of μ opioid receptors is specific to cell type, localization and developmental stage (Pasternak and Pan, 2013). There are several regions in the CNS associated with high levels of µ opioid receptor mRNA, such as the periaquaeductal gray, locus coeruleus and the raphe magnus. Among the somatosensory regions, μ opioid receptor mRNA is abundant in the dorsal horn and dorsal root ganglia, the spinal trigeminal, ambiguus and tractus solitarii nuclei, and the thalamus (Pasternak and Pan, 2013). Hwang and colleagues have shown that epigenetic factors mediate the expression of μ opioid receptors according to developmental stage. Specifically, P19 embryonic carcinoma cells showed hypermethylation in the proximal promoter region of the gene encoding for this receptor, associated with gene silencing. In contrast, the promoter was unmethylated, and the gene stably expressed when differentiation into mature neuronal/astrocyte cells was induced with retinoic acid (Hwang et al., 2007). At the same time, differentiation led to decreased methyl-CpG-binding protein 2 (MeCP2) binding in the promoter region of the gene for μ opioid receptors (Hwang et al., 2007). MeCP2 binds to methylated DNA regions and causes histone deacetylation, with subsequent compacting of the histone complex and decreased transcription (Nan et al., 1998). Therefore, both DNA methylation and histone modification contribute to regulating µ-opioid receptor expression.

Clinically, μ opioid receptor agonists, with morphine as their prototype compound, have remained the most relevant subgroup of opiates. Opiates are indispensable in modern perioperative medicine, but recent research has shown that multimodal pain treatment using several different classes of drugs can decrease the amount of opiates administered, and adverse effects (McCarthy *et al.*, 2010). The most important opiate side effects are nausea, vomiting, and bowel paralysis (Sharma and Jamal, 2013), suppression of the respiratory drive (Naso-Kaspar *et al.*, 2013), and opioid-induced hyperalgesia (OIH) (Bekhit, 2010).

Hyperalgesia induced by opiates is a clinically relevant phenomenon seen after both short- and long-term administration of opiates. The risk of inducing chronic hyperalgesia seems to vary with the opiate used, and is assumed to be highest for remifentanil (van Gulik et al., 2012), while the partial µ receptor agonist and κ receptor antagonist, buprenorphine has even been associated with an antihyperalgesic effect (Pergolizzi et al., 2010). OIH is prevented by the administration of the NMDA receptor antagonist, ketamine (Laulin et al., 2002). Mechanisms conjectured in animal and human studies include NMDA receptor activation, cholecystokinin, NO and neuroinflammation, with the end result of a change in gene expression, and adaptations in nociceptive systems (Bekhit, 2010). Liang and colleagues were able to demonstrate in mice that inhibition of the histone acetyltransferase in the presence of morphine reduced signs of OIH,



while histone deacetylase inhibition exaggerated OIH (Liang et al., 2013). In a recent clinical study, methylation analysis was undertaken in patients who had undergone methadone substitution following heroin addiction, patients receiving potent opiates for chronic pain, and age-matched controls. In the leukocytes of patients treated with various opioids, there was a global increase of DNA methylation at (LINE-1) and also a specific increase of methylation at the µ opioid receptor 1. Interestingly, a positive correlation was described between the methylation levels and pain intensity in patients treated with opiates, but not in patients in control groups receiving other pain medications (Doehring et al., 2013). It should be kept in mind, however, that the opiates constitute a heterogeneous group of drugs with distinct physicochemical properties, and the epigenetic effects of many opiates widely used perioperatively, such as tramadol (Scott and Perry, 2000), have not been investigated.

In conclusion, opiates remain of paramount importance in perioperative pain treatment. Both the developmental expression of the μ opioid receptor, and pathological hyperalgesia are mediated, at least in part, by epigenetic mechanisms. In general, opiates seem to increase global DNA methylation levels. This is in contrast to another group of perioperative drugs – the local anaesthetics.

Local anaesthetics

Just like the opiates, local anaesthetics are integral to perioperative medicine. They are a heterogeneous group of compounds characterized by the ability to block voltage-gated sodium channels, leading to distal anaesthesia and/or analgesia. For more than a hundred years, local anaesthetics have been used to provide neuraxial or peripheral anaesthesia and analgesia. However, recent research has shown that a substantial share of the effects of regional anaesthesia is due to systemic effects secondary to resorption from the site of injection (Lirk and Hollmann, 2013), and i.v. administration of, for example, lidocaine can replicate some effects of epidural anaesthesia (Herroeder et al., 2007). Specifically, some trials with local anaesthetics have demonstrated antinociceptive properties (Koppert et al., 2004), anti-inflammatory effects suppressing virtually every step of the inflammatory cascade (Hollmann and Durieux, 2000) and a reduction in perioperative hypercoagulability without inducing a clinically relevant bleeding risk (Hollmann et al., 2001).

Two studies have demonstrated the epigenetic effects of local anaesthetics. In breast cancer cells *in vitro*, the prototype local anaesthetic, lidocaine, induced demethylation of DNA as shown by MethyLight assays in a time- and concentration-dependent manner (Lirk *et al.*, 2012). These effects were more prominent in BT-20 oestrogen receptor-negative cells than in MCF-7 oestrogen-receptor positive cells, as BT-20 cells featured baseline methylation levels 100-fold higher than MCF-7 cells, and were observed at clinically relevant concentrations as seen during epidural anaesthesia (Fukuda *et al.*, 2003) or i.v. administration of lidocaine (Sun *et al.*, 2012). In a subsequent study, it was found that these effects could also be observed with another local anaesthetic ropivacaine, but not with bupivacaine. Also, there were no supra-additive

effects when treatment was combined with the classic demethylating drug, 5-aza-2'-deoxycytidine (Lirk *et al.*, 2014). The mechanism of demethylation remains unclear for contemporary amide-type local anaesthetics such as lido-caine, but research on ester-type local anaesthetics, a related group of drugs, showed that procaine had an inhibitory effect on DNA methyltransferase 1, the enzyme responsible for the maintenance methyltransferase activity of the cell (Castellano *et al.*, 2008).

These results are of potential interest for the development of chronic pain and cancer medicine. While opiates lead to increased methylation correlating with increased levels of pain suffering (Doehring et al., 2011), local anaesthetics have the potential to induce global DNA demethylation (Lirk et al., 2012), and may thereby protect against the development of hyperalgesia. Clinically, some studies have described an antinociceptive effect of local anaesthetics (Koppert et al., 2004), but others have found no clinically relevant effects (Herroeder et al., 2007; Martin et al., 2008). Whether perioperative anaesthetic technique can influence cancer recurrence after surgery remains fiercely debated, and most long-term studies are projected to last until the end of the decade because of long follow-up periods (Snyder and Greenberg, 2010). In this context, the therapeutic-demethylating levels of local anaesthetics in the perioperative setting may be of potential benefit, but the direct clinical relevance remains to be tested.

In conclusion, local anaesthetics are frequently administered perioperatively, and in contrast to opiates, they have a global DNA demethylating effect. This effect is of potential interest in the development of chronic pain and perioperative cancer medicine.

Perioperative inflammation

As part of an ancient evolutionary response, surgery is accompanied by a complex reaction referred to as 'surgical stress response'. This response shares substantial similarities with traumatic or burn injury, and is primarily mediated by the hypothalamic-pituitary axis leading to the release of stress hormones such as cortisol, growth hormone and vasopressin (Finnerty et al., 2013). It encompasses a wide variety of inflammatory, immune, hormonal and genomic responses (Finnerty et al., 2013). The expression of inflammatory genes is regulated by epigenetic factors such as histone acetylation (Barnes, 2013). This type of histone modification can be recognized by the highly conserved bromodomain motif of several epigenetic proteins. Low MW inhibitors of these bromodomains, such as JQ1, can inhibit the transcription of cytokines from mouse macrophages and protect against lipopolysaccharide-induced death (Belkina et al., 2013). Not only histone acetylation, but also histone methylation regulates the expression of inflammatory genes. Thus, histone methylation can reduce TNF release from human macrophages in vitro. The classic local anaesthetic, lidocaine, which has been shown to demethylate DNA, has very strong anti-inflammatory properties (Herroeder et al., 2007), but any causal role by epigenetic regulatory mechanisms has not yet been directly investigated.



Even though no direct investigation has been carried out, the potential influence of epigenetics on the inflammatory response that follows surgery seems a promising research area.

Wound healing

The repair of surgical wounds is a complex adaptive process where initial fibrogenesis leads to the formation of a scar, which is subsequently replaced by healthy and appropriate cells (Mann and Mann, 2013). Using bone healing as experimental paradigm, Bais and colleagues demonstrated the complex temporal sequence of transcriptome changes during healing, involving more than 300 genes (Bais et al., 2009). This confirms previous investigations that demonstrate the involvement of epigenetic mechanisms in the induction of wound healing (Shaw and Martin, 2009). Also, the migration of human cells, critical for wound healing, is regulated by epigenetic mechanisms, including the histone deacetylase 7 (Zheng et al., 2012). Epigenetics provides a basis to explain how certain disease states can negatively influence wound healing. For example, long-standing diabetes can impair physiological wound healing by decreasing the expression of the enzyme dicer, a pro-angiogenetic molecule, in a mouse model of type 2 diabetes. Similarly, the proliferation of keratinocytes, key to re-epithelialization of wound surfaces, is regulated by epigenetic factors (Rafehi et al., 2011). Moreover, alcoholics have a higher risk of nonunion after bone fractures, and this has been linked to epigenetic changes affecting cell migration, proliferation and differentiation (Sampson et al., 2011). Zeybel and co-workers investigated whether these epigenetic traits might be handed down through generations, and in a rat model of liver damage, they found that the fibrogenic component of liver damage was still alleviated in the first- and secondgeneration offspring after ancestral chronic liver injury induced by carbon tetrachloride (Zeybel et al., 2012). Lastly, epigenetics provides a therapeutic approach to regulating wound healing, for example by selectively targeting the promoter activity of relevant genes using oligodeoxynucleotides (Cutroneo and Chiu, 2000).

In conclusion, epigenetic mechanisms govern physiological wound healing, while the epigenetic side effects of diseases such as diabetes and alcoholism may explain erroneous wound healing. Therefore, epigenetics has been suggested as a therapeutic strategy to control wound healing.

Perioperative epigenetics and tumour recurrence

The perioperative period has long been recognized as a vulnerable time frame during which tumour progression and metastasis is often accelerated (Coffey *et al.*, 2003). Indeed, some small and retrospective trials have demonstrated improved patient survival and disease-free intervals when regional anaesthesia was used for cancer surgery (Schlagenhauff *et al.*, 2000; Exadaktylos *et al.*, 2006). Several experimental studies support this assumption. In a landmark experiment, growth in a tumour cell line was halted *in vitro* when incubated with serum taken from patients undergoing surgery under regional anaesthesia, but no effect was observed when serum from patients given general anaesthesia was applied (Deegan *et al.*, 2009). At this moment, several large-scale, multicentre, randomized and controlled trials are under way to investigate any beneficial effects of regional anaesthesia in tumour surgery, but they are projected to last until the end of the decade because of long follow-up times (Snyder and Greenberg, 2010).

The potential mechanisms for anti-metastatic effects from regional anaesthesia are either direct or indirect. Direct effects could include the interference of local anaesthetics with tumour-promoting pathways (Votta-Velis et al., 2013), direct toxic effects when used for local infiltration (Schlagenhauff et al., 2000) and changes in the epigenetic signature of tumour cells (Lirk et al., 2012). Indirect effects could result from reduction of the perioperative stress response and the preservation of the immune response (Dong et al., 2012). As many of these effects can be duplicated using i.v. local anaesthetics (Herroeder et al., 2007), a new avenue of research will be the employment of systemic local anaesthetics in the perioperative period of tumour surgery. Notably, local anaesthetics, at clinically relevant doses, demethylate DNA in breast cancer cells in vitro, opening up the possibility that epigenetic mechanisms could potentially explain why some anaesthetic techniques may have an anti-metastatic effect.

In conclusion, there is limited evidence suggesting that perioperative interventions may influence the recurrence of cancer after surgery. Epigenetics has been suggested as one potential pathway of interest.

Future directions

Research is needed to define whether perioperative surgical and anaesthetic interventions have the potential to alter epigenetic signatures and thereby potentially influence longterm patient outcome. In particular, despite the involvement of pain pathways, the 'epigenetic potentials' of widely used opiates, of NMDA antagonists and of non-opioid pain medications are unknown. In line with corresponding genetic research, epigenetic factors that render individual patients susceptible to development of chronic pain should be identified, and the concept of pain vulnerability (Denk et al., 2014) should be further developed and refined. The ultimate aim would be to predict, based on the type of intervention (Kehlet et al., 2006), patient history (Kalkman et al., 2003) and genetic and epigenetic factors (Denk et al., 2014), which patients are at risk of experiencing acute and chronic pain, and to tailor the anaesthetic and surgical plan accordingly, allocating analgesic resources to patients who need it most. For example, this may influence the choice of administering regional anaesthesia or multimodal analgesia, or both (Lirk and Hollmann, 2013).

Epigenetic factors in the perioperative period are only beginning to be understood and have major therapeutic and diagnostic potential.



Acknowledgement

The authors gratefully acknowledge the expert language editing assistance of Mrs Senay Boztas, Amsterdam, The Netherlands.

Author contributions

P.L. wrote the manuscript and serves as the corresponding author. H.F. assisted in writing the manuscript and approved the final version. N.C.W. assisted in writing the manuscript and approved the final version. M.W.H. wrote the manuscript and is the senior author.

Conflict of interest

None declared.

References

Alexander SPH, Benson HE, Faccenda E, Pawson AJ, Sharman JL, Spedding M *et al.* (2013a). The Concise Guide to PHARMACOLOGY 2013/14: G Protein-Coupled Receptors. Br J Pharmacol 170: 1459–1581.

Alexander SPH, Benson HE, Faccenda E, Pawson AJ, Sharman JL, Spedding M *et al.* (2013b). The Concise Guide to PHARMACOLOGY 2013/14: Enzymes. Br J Pharmacol 170: 1797–1867.

Alexander SPH, Benson HE, Faccenda E, Pawson AJ, Sharman JL, Catterall WA *et al.* (2013c). The Concise Guide to PHARMACOLOGY 2013/14: Ion Channels. Br J Pharmacol 170: 1607–1651.

Bain CR, Shaw AD (2012). Genetics and epigenetics in perioperative medicine. Curr Opin Crit Care 18: 548–554.

Bais M, McLean J, Sebastiani P, Young M, Wigner N, Smith T *et al.* (2009). Transcriptional analysis of fracture healing and the induction of embryonic stem cell-related genes. PLoS ONE 4: e5393.

Barnes PJ (2013). New anti-inflammatory targets for chronic obstructive pulmonary disease. Nat Rev Drug Discov 12: 543–559.

Bekhit MH (2010). Opioid-induced hyperalgesia and tolerance. Am J Ther 17: 498–510.

Belkina AC, Nikolajczyk BS, Denis GV (2013). BET protein function is required for inflammation: Brd2 genetic disruption and BET inhibitor JQ1 impair mouse macrophage inflammatory responses. J Immunol 190: 3670–3678.

Bell JT, Loomis AK, Butcher LM, Gao F, Zhang B, Hyde CL *et al.* (2014). Differential methylation of the TRPA1 promoter in pain sensitivity. Nat Commun 5: 2978.

Buchheit T, Van de Ven T, Shaw A (2012). Epigenetics and the transition from acute to chronic pain. Pain Med 13: 1474–1490.

Castellano S, Kuck D, Sala M, Novellino E, Lyko F, Sbardella G (2008). Constrained analogues of procaine as novel small molecule inhibitors of DNA methyltransferase-1. J Med Chem 51: 2321–2325.

Chiou LC, Fan SH, Chuang KC, Liao YY, Lee SZ (2004). Pharmacological characterization of nociceptin/orphanin FQ receptors, a novel opioid receptor family, in the midbrain periaqueductal gray. Ann N Y Acad Sci 1025: 398–403.

Coffey JC, Wang JH, Smith MJ, Bouchier-Hayes D, Cotter TG, Redmond HP (2003). Excisional surgery for cancer cure: therapy at a cost. Lancet Oncol 4: 760–768.

Correll DJ, Vlassakov KV, Kissin I (2014). No evidence of real progress in treatment of acute pain, 1993–2012: scientometric analysis. J Pain Res 7: 199–210.

Costigan M, Belfer I, Griffin RS, Dai F, Barrett LB, Coppola G *et al.* (2010). Multiple chronic pain states are associated with a common amino acid-changing allele in KCNS1. Brain 133: 2519–2527.

Cutroneo KR, Chiu JF (2000). Comparison and evaluation of gene therapy and epigenetic approaches for wound healing. Wound Repair Regen 8: 494–502.

Deegan CA, Murray D, Doran P, Ecimovic P, Moriarty DC, Buggy DJ (2009). Effect of anaesthetic technique on oestrogen receptor-negative breast cancer cell function *in vitro*. Br J Anaesth 103: 685–690.

Denk F, McMahon SB, Tracey I (2014). Pain vulnerability: a neurobiological perspective. Nat Neurosci 17: 192–200.

Doehring A, Geisslinger G, Lotsch J (2011). Epigenetics in pain and analgesia: an imminent research field. Eur J Pain 15: 11–16.

Doehring A, Oertel BG, Sittl R, Lotsch J (2013). Chronic opioid use is associated with increased DNA methylation correlating with increased clinical pain. Pain 154: 15–23.

Dong H, Zhang Y, Xi H (2012). The effects of epidural anaesthesia and analgesia on natural killer cell cytotoxicity and cytokine response in patients with epithelial ovarian cancer undergoing radical resection. J Int Med Res 40: 1822–1829.

Exadaktylos AK, Buggy DJ, Moriarty DC, Mascha E, Sessler DI (2006). Can anesthetic technique for primary breast cancer surgery affect recurrence or metastasis? Anesthesiology 105: 660–664.

Finnerty CC, Mabvuure NT, Ali A, Kozar RA, Herndon DN (2013). The surgically induced stress response. JPEN J Parenter Enteral Nutr 37: 21S–29S.

Fukuda T, Kakiuchi Y, Miyabe M, Kihara S, Kohda Y, Toyooka H (2003). Free lidocaine concentrations during continuous epidural anesthesia in geriatric patients. Reg Anesth Pain Med 28: 215–220.

van Gulik L, Ahlers SJ, van de Garde EM, Bruins P, van Boven WJ, Tibboel D *et al.* (2012). Remifentanil during cardiac surgery is associated with chronic thoracic pain 1 yr after sternotomy. Br J Anaesth 109: 616–622.

Heijmans BT, Tobi EW, Stein AD, Putter H, Blauw GJ, Susser ES *et al.* (2008). Persistent epigenetic differences associated with prenatal exposure to famine in humans. Proc Natl Acad Sci U S A 105: 17046–17049.

Herroeder S, Pecher S, Schonherr ME, Kaulitz G, Hahnenkamp K, Friess H *et al.* (2007). Systemic lidocaine shortens length of hospital stay after colorectal surgery: a double-blinded, randomized, placebo-controlled trial. Ann Surg 246: 192–200.

Hollmann MW, Durieux ME (2000). Local anesthetics and the inflammatory response: a new therapeutic indication? Anesthesiology 93: 858–875.

Hollmann MW, Wieczorek KS, Smart M, Durieux ME (2001). Epidural anesthesia prevents hypercoagulation in patients undergoing major orthopedic surgery. Reg Anesth Pain Med 26: 215–222.



Hwang CK, Song KY, Kim CS, Choi HS, Guo XH, Law PY *et al*. (2007). Evidence of endogenous mu opioid receptor regulation by epigenetic control of the promoters. Mol Cell Biol 27: 4720–4736.

Imai S, Ikegami D, Yamashita A, Shimizu T, Narita M, Niikura K *et al.* (2013). Epigenetic transcriptional activation of monocyte chemotactic protein 3 contributes to long-lasting neuropathic pain. Brain 136: 828–843.

Julius D (2013). TRP channels and pain. Annu Rev Cell Dev Biol 29: 355–384.

Kalkman CJ, Visser K, Moen J, Bonsel GJ, Grobbee DE, Moons KG (2003). Preoperative prediction of severe postoperative pain. Pain 105: 415–423.

Kehlet H, Jensen TS, Woolf CJ (2006). Persistent postsurgical pain: risk factors and prevention. Lancet 367: 1618–1625.

Kern U, Busch V, Rockland M, Kohl M, Birklein F (2009). Prevalence and risk factors of phantom limb pain and phantom limb sensations in Germany. A nationwide field survey. Schmerz 23: 479–488.

Koppert W, Weigand M, Neumann F, Sittl R, Schuettler J, Schmelz M *et al.* (2004). Perioperative intravenous lidocaine has preventive effects on postoperative pain and morphine consumption after major abdominal surgery. Anesth Analg 98: 1050–1055.

Kruidenier L, Chung CW, Cheng Z, Liddle J, Che K, Joberty G *et al.* (2012). A selective jumonji H3K27 demethylase inhibitor modulates the proinflammatory macrophage response. Nature 488: 404–408.

Laulin JP, Maurette P, Corcuff JB, Rivat C, Chauvin M, Simonnet G (2002). The role of ketamine in preventing fentanyl-induced hyperalgesia and subsequent acute morphine tolerance. Anesth Analg 94: 1263–1269.

Liang DY, Li X, Clark JD (2013). Epigenetic regulation of opioid-induced hyperalgesia, dependence, and tolerance in mice. J Pain 14: 36–47.

Lirk P, Hollmann M (2013). Outcome after regional anesthesia: weighing risks and benefits. Minerva Anestesiol 80: 610–618.

Lirk P, Berger R, Hollmann MW, Fiegl H (2012). Lidocaine timeand dose-dependently demethylates deoxyribonucleic acid in breast cancer cell lines *in vitro*. Br J Anaesth 109: 200–207.

Lirk P, Hollmann M, Fleischer M, Hauck-Weber NC, Fiegl H (2014). Lignocaine and ropivacaine, but not bupivacaine, demethylate deoxyribonucleic acid in breast cancer cells *in vitro*. Br J Anaesth 113 (Suppl. 1): i32–i38.

Mann J, Mann DA (2013). Epigenetic regulation of wound healing and fibrosis. Curr Opin Rheumatol 25: 101–107.

Martin DS, Khosravi M, Grocott MP, Mythen MG (2010). Concepts in hypoxia reborn. Crit Care 14: 315.

Martin F, Cherif K, Gentili ME, Enel D, Abe E, Alvarez JC *et al.* (2008). Lack of impact of intravenous lidocaine on analgesia, functional recovery, and nociceptive pain threshold after total hip arthroplasty. Anesthesiology 109: 118–123.

McCarthy GC, Megalla SA, Habib AS (2010). Impact of intravenous lidocaine infusion on postoperative analgesia and recovery from surgery: a systematic review of randomized controlled trials. Drugs 70: 1149–1163.

Naguib M, Bie B, Ting AH (2012). Fundamental concepts of epigenetics for consideration in anesthesiology. Curr Opin Anaesthesiol 25: 434–443.

Nan X, Ng HH, Johnson CA, Laherty CD, Turner BM, Eisenman RN *et al.* (1998). Transcriptional repression by the methyl-CpG-binding protein MeCP2 involves a histone deacetylase complex. Nature 393: 386–389.

Naso-Kaspar CK, Herndon GW, Wyman JF, Felo JA, Lavins ES, Gilson TP (2013). 'Lingering' opiate deaths? Concentration of opiates in medulla and femoral blood. J Anal Toxicol 37: 507–511.

Navada SC, Steinmann J, Lubbert M, Silverman LR (2014). Clinical development of demethylating agents in hematology. J Clin Invest 124: 40–46.

Norbury TA, MacGregor AJ, Urwin J, Spector TD, McMahon SB (2007). Heritability of responses to painful stimuli in women: a classical twin study. Brain 130: 3041–3049.

Nyman T, Mulder M, Iliadou A, Svartengren M, Wiktorin C (2011). High heritability for concurrent low back and neck-shoulder pain: a study of twins. Spine (Phila Pa 1976) 36: E1469–E1476.

Pasternak GW (2004). Multiple opiate receptors: deja vu all over again. Neuropharmacology 47 (Suppl. 1): 312–323.

Pasternak GW, Pan YX (2013). Mu opioids and their receptors: evolution of a concept. Pharmacol Rev 65: 1257–1317.

Pawson AJ, Sharman JL, Benson HE, Faccenda E, Alexander SP, Buneman OP *et al.*; NC-IUPHAR (2014). The IUPHAR/BPS Guide to PHARMACOLOGY: an expert-driven knowledge base of drug targets and their ligands. Nucl Acids Res 42 (Database Issue): D1098–D1106.

Pergolizzi J, Aloisi AM, Dahan A, Filitz J, Langford R, Likar R *et al.* (2010). Current knowledge of buprenorphine and its unique pharmacological profile. Pain Pract 10: 428–450.

Portela A, Esteller M (2010). Epigenetic modifications and human disease. Nat Biotechnol 28: 1057–1068.

Rafehi H, El-Osta A, Karagiannis TC (2011). Genetic and epigenetic events in diabetic wound healing. Int Wound J 8: 12–21.

Reiber GE, McFarland LV, Hubbard S, Maynard C, Blough DK, Gambel JM *et al.* (2010). Servicemembers and veterans with major traumatic limb loss from Vietnam war and OIF/OEF conflicts: survey methods, participants, and summary findings. J Rehabil Res Dev 47: 275–297.

Richardson C, Glenn S, Nurmikko T, Horgan M (2006). Incidence of phantom phenomena including phantom limb pain 6 months after major lower limb amputation in patients with peripheral vascular disease. Clin J Pain 22: 353–358.

Sampson HW, Chaput CD, Brannen J, Probe RA, Guleria RS, Pan J *et al.* (2011). Alcohol induced epigenetic perturbations during the inflammatory stage of fracture healing. Exp Biol Med (Maywood) 236: 1389–1401.

Schlagenhauff B, Ellwanger U, Breuninger H, Stroebel W, Rassner G, Garbe C (2000). Prognostic impact of the type of anaesthesia used during the excision of primary cutaneous melanoma. Melanoma Res 10: 165–169.

Scott LJ, Perry CM (2000). Tramadol: a review of its use in perioperative pain. Drugs 60: 139–176.

Sharma A, Jamal MM (2013). Opioid induced bowel disease: a twenty-first century physicians' dilemma. Considering pathophysiology and treatment strategies. Curr Gastroenterol Rep 15: 334.

Shaw T, Martin P (2009). Epigenetic reprogramming during wound healing: loss of polycomb-mediated silencing may enable upregulation of repair genes. EMBO Rep 10: 881–886.

Perioperative epigenetics



Snyder GL, Greenberg S (2010). Effect of anaesthetic technique and other perioperative factors on cancer recurrence. Br J Anaesth 105: 106–115.

Sun Y, Li T, Wang N, Yun Y, Gan TJ (2012). Perioperative systemic lidocaine for postoperative analgesia and recovery after abdominal surgery: a meta-analysis of randomized controlled trials. Dis Colon Rectum 55: 1183–1194.

Tajerian M, Alvarado S, Millecamps M, Vachon P, Crosby C, Bushnell MC *et al.* (2013). Peripheral nerve injury is associated with chronic, reversible changes in global DNA methylation in the mouse prefrontal cortex. PLoS ONE 8: e55259.

Teschendorff AE, Menon U, Gentry-Maharaj A, Ramus SJ, Weisenberger DJ, Shen H *et al.* (2010). Age-dependent DNA methylation of genes that are suppressed in stem cells is a hallmark of cancer. Genome Res 20: 440–446.

Votta-Velis EG, Piegeler T, Minshall RD, Aguirre J, Beck-Schimmer B, Schwartz DE *et al.* (2013). Regional anaesthesia and cancer

metastases: the implication of local anaesthetics. Acta Anaesthesiol Scand 57: 1211–1229.

van den Wijngaard RM, Stanisor OI, van Diest SA, Welting O, Wouters MM, Cailotto C *et al.* (2013). Susceptibility to stress induced visceral hypersensitivity in maternally separated rats is transferred across generations. Neurogastroenterol Motil 25: e780–e790.

Zeybel M, Hardy T, Wong YK, Mathers JC, Fox CR, Gackowska A *et al.* (2012). Multigenerational epigenetic adaptation of the hepatic wound-healing response. Nat Med 18: 1369–1377.

Zhang Z, Cai YQ, Zou F, Bie B, Pan ZZ (2011). Epigenetic suppression of GAD65 expression mediates persistent pain. Nat Med 17: 1448–1455.

Zheng C, Yu Z, Zhou Y, Tao L, Pang Y, Chen T *et al.* (2012). Live cell imaging analysis of the epigenetic regulation of the human endothelial cell migration at single-cell resolution. Lab Chip 12: 3063–3072.