

Nutraceuticals against Neurodegeneration: A Mechanistic Insight

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Abstract: The mechanisms underlying neurodegenerative disorders are complex and multifactorial; however, accumulating evidences suggest few common shared pathways.

These common pathways include mitochondrial dysfunction, intracellular Ca^{2+} overload, oxidative stress and inflammation. Often multiple pathways co-exist, and therefore limit the benefits of therapeutic interventions. Nutraceuticals have recently gained importance owing to their multifaceted effects. These food-based approaches are believed to target multiple pathways in a slow but more physiological manner without causing severe adverse effects. Available information strongly supports the notion that apart from preventing the onset of neuronal damage, nutraceuticals can potentially attenuate the continued progression of neuronal destruction. In this article, we i) review the common pathways involved in the pathogenesis of the toxicants-induced neurotoxicity and neurodegenerative disorders with special emphasis on Alzheimer's disease (AD), Parkinson's disease (PD), Huntington's disease (HD), Multiple sclerosis (MS) and Amyotrophic lateral sclerosis (ALS), and ii) summarize current research advancements on the effects of nutraceuticals against these detrimental pathways.

Keywords: Neurodegeneration, mitochondria, calcium, oxidative stress, inflammation, nutraceuticals.

Received: June 18, 2015

Revised: August 17, 2015

Accepted: January 01, 2016

1. INTRODUCTION

Neurodegenerative diseases are one of the most debilitating conditions and usually associated with mutated genes, accumulation of abnormal proteins, increased reactive oxygen species (ROS) or destruction of the neurons in a specific part of the brain [1-5]. Improved experimental models, modern research techniques and research in the field of neuroscience have advanced our understanding about the unique mechanisms of the onset and expansion of the neurodegeneration and toxicants-induced neurotoxicity. Among several recognized mechanisms, the damage to neuronal mitochondria, intracellular Ca^{2+} overload, uncontrolled generation of ROS, sustained inflammatory condition or any combination of these are the common busy high-roads for neurotoxicity [6-14]. Increased oxidative stress and mild chronic inflammation are the host of multiple disorders, including neuronal disorders. On the other hand, Ca^{2+} homeostasis across the membrane (cellular, endoplasmic reticulum (ER) and mitochondrial) is critical for a variety of

cellular physiological functions. The multiple order of calcium concentration [Ca^{2+}] gradient across the membrane is precisely regulated through several specialized mechanisms. Any event that perturbs these mechanisms leads to the persistent increase in the intracellular and intra-organelle (ER and mitochondria) [Ca^{2+}], which has been linked to the pathogenesis of several neurodegenerative disorders [15-17]. Among these mechanisms, mitochondrial damage has gained particular attention, as perturbation of its function triggers activation of neurotoxic pathways such as deregulation of Ca^{2+} homeostasis, generation of ROS from the electron transport chain (ETC) and impaired cellular energy production [18]. Although, the causative factor for the onset of these signaling may differ, the progression of toxicants-induced neuronal damage and neurodegenerative disorders largely involves the same shared mechanisms, reinforcing the importance of these pathways as common targets for the intervention strategies. Advancement in our understanding of the mechanisms underlying the neuronal damage has made it possible to design interventions that can potentially delay the onset and/or prevent further expansion of the unabated neurodegeneration [19, 20].

Utilizing the knowledge of the etiology of the neurodegenerative diseases, target-based therapies such as neurotransmitter modulators, direct receptor agonists/antagonists, second messenger modulators, stem cells-based therapies, hormone replacement therapy, neurotrophic factors as well

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as regulators of the mRNA synthesis and its translation into disease-causing mutant proteins have been developed [21-28]. Although, these strategies are great tools to mitigate the neurodegenerative processes, such treatments are often associated with adverse effects and long-term unknown consequences [22, 25, 29, 30]. Neural transplantation and stem cells-based therapies are emerging with a great hope to fight against the range of neurodegenerative diseases [31-33]. Despite unprecedented advancement in these fields of research, the therapeutic applications of such novel tools are still far from reaching the clinics. So, there is a need to identify a safer alternative treatment option that can be employed over long duration to fight against the debilitating neuronal conditions. Nutraceuticals, which can simultaneously act on multiple targets to improve the overall neuronal health, appear to be an attractive option [34-39]. In the present article, we have reviewed mitochondrial dysfunction, intracellular Ca^{2+} overload, oxidative stress and inflammation as common pathways involved in the neuronal damage and evaluated current research advancements on the effects of some selected nutraceuticals on these pathways.

2. MECHANISMS OF NEURONAL DAMAGE

2.1. Mitotoxicity-mediated Neuronal Damage

Mitochondrial damage underlies the pathogenesis of several types of toxicants-induced acute neurotoxicity and neurodegenerative diseases [40-48]. Unlike other organelles, the regulation of mitochondrial functions is under the control of two genomes: nuclear DNA (nDNA) and mitochondrial DNA (mtDNA). Mutations in either of these genomes can result in the mitotoxicity-mediated neurodegeneration [49]. Mitochondrial dysfunction could affect distinct compartments of a single neuron or a specific region of the brain. Widely studied glutamate neurotoxicity by Ca^{2+} influx is also primarily attributed to the mitochondrial permeability transition pore (MPT) [40, 45]. The neuro-toxicant, 1-methyl-4-phenylpyridinium (MPP^+), as generated from the mono amine oxidase (MAO)-catalyzed oxidation of 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP) within the brain, is concentrated inside the mitochondria of the dopaminergic cells and inhibits Complex I of the respiratory electron transport chain (ETC), which contributes to the development of Parkinson's disease (PD) [50, 51]. Poly ADP ribose polymerase (PARP) activation-associated neuronal cell death (both apoptotic and necrotic forms) after cerebral ischemia is mediated by cyclophilin D (CypD)-dependent MPT [50, 51]. Neurotoxicity that occurs with various chemotherapeutic agents such as taxanes [52], bortezomib [53] and platinum compounds [54] is also associated with the mitochondrial dysfunction.

Pathophysiology of several neurodegenerative diseases such as Alzheimer's disease (AD), PD, Huntington's disease (HD), multiple sclerosis (MS) and amyotrophic lateral sclerosis (ALS) largely involves damaged mitochondria [55-59]. Several mitochondrial-dependent molecular mechanisms such as inhibition of mitochondrial ETC's complexes, generation of ROS, perturbations of mitochondrial clearance mechanisms, altered functioning of the enzymes involved in tricarboxylic acid cycle, impairment of mitochondrial dynamics, and interference with the timely supply of the

functional mitochondria to the specific sites within a neuron could contribute to the pathogenesis of neuronal injury and neurodegenerative diseases [40, 41, 43-46]. Defective ETC (Complex I) in the substantia-nigra has been considered as a central cause of the sporadic form of PD [50, 51, 57, 60, 61]. Mitochondrial abnormalities also underlie the neuronal damage in AD [55, 62] and ALS [59, 63, 64]. Neuronal degeneration in MS and in experimental autoimmune encephalomyelitis (EAE) is also attributed to the abnormal activities of Complexes I and IV of the ETC as well as the loss of mitochondrial membrane potential ($\Delta\Psi_m$) [65, 66]. Evidence from pre-clinical and clinical studies suggested the role of mitotoxicity in the pathogenesis of neuronal degeneration in HD [56, 67-70]. Mutant huntingtin, the gene responsible for the development of HD, is reported to directly impair mitochondrial functions [71]. Defective mitochondrial Complex II, III and IV have been found in the postmortem tissue from HD patients [72, 73]. Several studies have concluded a positive relationship between mtDNA fragmentation in striatal and cortical neurons and the development of HD [74].

Collectively, these findings suggest that mitochondrial damage is the primary event observed in toxicants-induced neurotoxicity as well as various neurodegenerative diseases. Owing to complex interplay between mitotoxicity, oxidative stress and sustained inflammatory status, it has always remained unclear if mitochondrial damage is the cause or consequence of neuronal damage. Several lines of evidence suggest that mitotoxicity-associated intracellular Ca^{2+} overload, oxidative stress and inflammation are the leading causes for the development of neurodegeneration [43]. The role of Ca^{2+} overload, oxidative stress and inflammation in the development of neurotoxicity and neurodegeneration has been discussed below.

2.2. Ca^{2+} Overload-mediated Neuronal Damage

Research about the role of Ca^{2+} ions in neurotoxicity and neurodegenerative diseases started in 1970s [75, 76]. In general, the $[\text{Ca}^{2+}]$ in extracellular space remains in millimolar range, while inside cells, it remains in micromolar range [75, 77-79]. This multiple order of $[\text{Ca}^{2+}]$ difference across plasma membrane under normal physiological conditions is attributed to the selective permeability of plasma membrane, the dynamic functions of the several adenosine triphosphate (ATP)-dependent Ca^{2+} -ATPase pumps and sodium/calcium exchange system that actively pumps Ca^{2+} ions outside the cells [17, 80, 81]. Moreover intracellular low level of $[\text{Ca}^{2+}]$ is maintained by the buffering functions of ER and mitochondria by virtue of their ability to store the Ca^{2+} ions [17, 47, 82]. Storage of Ca^{2+} inside the ER occurs after active transport of cytoplasmic Ca^{2+} through ATP-dependent pumps in the ER membrane [17]. However, uptake of cytoplasmic Ca^{2+} into mitochondria occurs through a membrane protein known as electrophoretic uniporter present in the mitochondrial outer membrane [47]. Cellular events that compromise integrity of the plasma membranes or the functions of ATP-dependent Ca^{2+} -ATPase pumps/uniporter system can potentially be responsible for rise in intracellular $[\text{Ca}^{2+}]$. In addition to these mechanisms, Ca^{2+} accumulation in neuronal cells can also occur through several other routes such as activation of

voltage-sensitive Ca^{2+} channels, receptor-operated Ca^{2+} channels (N-methyl-D-aspartate (NMDA)), ATP-dependent Ca^{2+} channel (P2j receptor), cyclic nucleotide-gated Ca^{2+} channels, Ca^{2+} channels coupled to G protein receptors and are sensitive to inositol 1,3,4,5 tetrakisphosphate [83]. Notably, the plasma membrane, ER and mitochondria can only handle rise in intracellular Ca^{2+} up to certain extent. Persistent rise in intracellular Ca^{2+} leads to disturbed ER and mitochondrial Ca^{2+} homeostasis, which has been linked to the pathogenesis of various neurodegenerative diseases [16, 17]. Excitotoxicity is also related to the influx of Ca^{2+} ions into the neurons [84]. Further, some pathological conditions such as cerebral ischemia and traumatic brain injury are also associated with an abrupt increase in intracellular $[\text{Ca}^{2+}]$ [85, 86]. Sudden rise in intracellular $[\text{Ca}^{2+}]$ is an immediate threat to the cell, as it triggers a cascade of neurotoxic events, including mitotoxicity and cessation of ATP synthesis, over-activation of several Ca^{2+} -dependent hydrolytic enzymes such as proteases, phospholipases, nucleases, nitric oxide synthase and phosphatases [87, 88]. These events lead to the onset of neurotoxicity, impaired neuronal functions and eventually death of the neuronal cells. This explains why the maintenance of the intracellular Ca^{2+} ions homeostasis is of crucial importance for the survival of the neuronal cells. Exposure to several neurotoxic chemicals result in the perturbed intracellular Ca^{2+} homeostasis, which eventually leads to the death of neuronal cells [89]. Increased intracellular $[\text{Ca}^{2+}]$ is not only implicated in the traumatic and toxicants-induced neurotoxicity, but several neurodegenerative diseases also have the same underlying mechanisms [90-93]. In AD, amyloid-beta ($\text{A}\beta$)-induced neuronal cell death is associated with the dysregulation of Ca^{2+} -dependent pathways [93-95]. Moreover, deregulation of the intracellular Ca^{2+} could explain the selective destruction of dopaminergic neurons in the PD [96]. Neuronal destabilization and injury in MS is due to the overactivation of the Ca^{2+} -dependent proteases, calpain and protein phosphatases [97]. Opening of the MPT due to mitochondrial Ca^{2+} overload and release of the Ca^{2+} ions from the ER by mutant huntingtin is considered as the mechanism of the neuronal death in HD [71, 98]. Collectively, this information indicates that the death of the neuronal cells in toxicants-induced neurotoxicity and neurodegenerative disease follow the common $[\text{Ca}^{2+}]$ -dependent mechanisms. However, further studies are needed to understand the downstream pathway that precisely orchestrate the expansion of high intracellular $[\text{Ca}^{2+}]$ -mediated death of neuronal cells in neurodegenerative diseases.

2.3. Oxidative Stress-mediated Neuronal Damage

Excessive production of ROS as a result of the imbalance among cellular biochemical processes gives rise to a condition known as oxidative stress [99]. ROS and reactive nitrogen species (RNS)-mediated damage to cellular macromolecules is involved in the pathogenesis of several neurodegenerative diseases [14, 100, 101]. Nervous system is particularly vulnerable to the oxidative stress due to several reasons such as: i) high oxygen consumption due to higher demand of energy (ATP), ii) relatively lower level of the endogenous antioxidants reserve, iii) abundance of polyunsaturated lipids, which are particularly susceptible to free radical attack, in the plasma membrane of large neuronal

cells, iv) high levels of redox-active transition metals, v) presence of excitatory amino acids and neurotransmitters, whose metabolism can produce ROS, and vi) almost complete dependency of the neuronal cells on the membrane-dependent dynamic synaptic transmission for the effective communication [102-107]. These characteristics make neuronal cells an easy target for the ROS- and RNS-mediated damage. Although, initial events after MPP^+ exposure is the inhibition of Complex I in ETC, subsequent oxidative stress is the underlying cause of the PD like condition [108]. The loss of dopaminergic and serotonergic nerve terminals in the methamphetamine (METH) and 3,4-methylenedioxymethamphetamine (MDMA) drug abusers has been attributed to the oxidative stress [109, 110]. Neuronal degeneration and $\text{A}\beta$ neurotoxicity in AD patients is associated with the oxidative damage to DNA, RNA, proteins and lipids [111-116]. Oxidative damage to DNA [117-119] and protein [120, 121] has been reported in the nigro-striatal region of the brain in PD. Oxidation of dopamine to a reactive 6-hydroxydopamine (6-OHDA) provides an important endogenous mechanism for the development of PD like condition [122-124]. Iron, a transition metal, overload has been recognized to play a key role in the Lewy body formation in PD [125]. Perturbed homeostasis of redox-active and inactive metals has been associated with the affected area in the brain of AD [113, 114] and PD [126-129]. Oxidative stress-induced mutations in the gene encoding for the ubiquitous Cu/Zn-superoxide dismutase (SOD-1) enzyme [130] and damage to the proteins, lipids and DNA [131-133] have been associated with the familial and sporadic forms of the ALS. Pathology of MS is no different when the detrimental effect of the oxidative stress is considered. ROS and RNS produced by the activated microglia and mononuclear cells play a major role in the pathogenesis of MS. Oxidative damage to nDNA, mtDNA and decreased level of endogenous antioxidants [134-136] have been linked to the demyelination [137] and axonal injury in MS and EAE [138]. Increased incidence of the oxidative DNA strand breaks and exacerbated lipofuscin, a pigment produced by the reaction of cellular amino compounds with the aldehydic products of the oxidative damage to tissue macromolecules, is associated with HD [139, 140]. Significant increase in the levels of 8-OH-dG in nDNA has been found in the postmortem tissue of HD cases [141]. All these evidences clearly indicate a causal relationship between the oxidative stress and neuronal cell death in neurodegenerative diseases and toxicants-induced neurotoxicity. This implies that interventional agents such as antioxidants, metal chelators and those able to boost the cellular endogenous antioxidants may offer a therapeutic strategy to prevent the continued progression and sometimes even initiation of the diseases.

2.4. Inflammation-mediated Neuronal Damage

Emerging evidence underscores an active role of neuro-inflammation in the pathophysiology of neurodegenerative diseases [142]. The initiation and subsequent propagation of the neuroinflammation appear to rely on the interaction between glia, immune cells and neurons. Immunologic privileges of brain owing to absence of physical connection between central nervous system and classical lymphatic

system has recently been challenged with identification of functional and classical lymphatic system in the brain [143] suggesting an important role in the pathogenesis of neuronal diseases. Macrophages are present in the brain near glia and microglia and play a fundamental role in the inflammation-mediated neurodegenerative disorders. In the disease state, activated microglia mediate neuronal injury through the production of pro-inflammatory factors such as cytokines and chemokines [144]. Production of pro-inflammatory cytokines and chemokines leads to the trans-endothelial migration of immune cells across the blood-brain-barrier [145]. Several mechanisms have been identified for the microglia-mediated phagocytic and cytotoxic actions that are responsible for neuronal damage. The first and foremost mechanism is phagocytic oxidase (PHOX)-mediated oxidative stress-induced neurotoxicity [145, 146]. Unlike healthy brain, when PHOX (normally expressed at high levels in microglia) is acutely stimulated by inflammatory condition, it causes oxidative stress by rapidly producing high levels of superoxides (O_2^-).

Inflammatory activation of PHOX also causes activation of microglia [146] resulting in the production of TNF α , IL-1 β and inducible NO Synthase (iNOS) [147, 148]. Thus, PHOX activation provides an important link between inflammation and oxidative stress [149]. Thus, inhibition of PHOX has been reported as a potential therapeutic strategy for several neuroinflammatory conditions. iNOS is expressed in microglia, astrocytes and is upregulated during inflammation [150, 151], resulting in increased NO production and thereby RNS leading to neuronal death through the inhibition of the mitochondrial cytochrome oxidase in the neurons [145, 152, 153]. Increased iNOS expression during inflammation may potentiate the neuronal death by hypoxic condition, as it can compete with cytochrome oxidase for the molecular oxygen [153, 154]. However, it should be noted that, activation of either NADPH oxidase (NOX/PHOX) or production of NO through iNOS expression alone is not sufficient to induce neurotoxicity, but their combined activation underlies the mechanism of inflammatory neurodegeneration [145, 155]. Apart from this, chemokines secreted by astrocytes play multiple roles in the pathology of a chronic inflammatory neurodegenerative disease such as AD [154, 156, 157]. Given the self-perpetuating cyclic nature of inflammatory processes in the progression of neurodegeneration [158], intervention with the natural compounds having strong anti-inflammatory activities may offer a safer way to prevent or at least reduce the progression of such devastating diseases.

2.5. Interplay between Intracellular Mitotoxicity, Ca^{2+} Accumulation, Oxidative Stress and Inflammation

Although, each mechanism discussed above can independently mediate neuronal damage, they are highly interlinked with each other and often co-exist. Every single mechanism can serve as an initiation point for the vicious cycle. But perturbation in Ca^{2+} homeostasis has immediate cytotoxic consequences Ca^{2+} overload, as observed during excitotoxicity, is sensed by the ER, mitochondria and membrane bound Ca^{2+} -transporter pumps. ER and mitochondria operate to re-establish intracellular Ca^{2+}

homeostasis. Continued uptake of Ca^{2+} by mitochondria through Ca^{2+} uniporter results in the dissipation of $\Delta\Psi_m$, hindering ATP synthase activity. Inhibition of ATP synthesis impairs the regulatory functions of ATP-dependent Ca^{2+} -ATPase pumps located in the membrane of ER and cell. This causes further accumulation of Ca^{2+} in the cytoplasm of the cells. Continuous overload of Ca^{2+} increases ROS production through interference with the ETC [159]. Ca^{2+} overload can activate constitutively expressed NOS in the neuronal and endothelial cells of the brain to produce excess amount of NO [160, 161]. Coproduction of NO and O_2^- leads to the formation of highly reactive NOO_2^- . Furthermore, NOO_2^- can itself augment its formation through the inactivation of Mn-SOD. ROS and RNS irreversibly bind with the respiratory complexes and inhibit ATP synthesis.

Excessive production of ROS and RNS can induce DNA strand break, which leads to the activation of DNA repair enzymes such as PARP. Activated PARP causes depletion of cellular NAD^+ , which severely compromises ATP synthesis. Thus, oxidative and nitrosative stress can indirectly lead to the accumulation of cytoplasmic Ca^{2+} by depriving the functions of ATP-dependent Ca^{2+} pumps. Furthermore, ROS and RNS can directly inactivate ATP-dependent Ca^{2+} pumps by reacting with the soft nucleophile, such as thiol group, and contribute towards the cytoplasmic overload of the Ca^{2+} ions. Moreover, mitochondrial dysfunction in the neurodegenerative diseases leads to the deregulation of the Ca^{2+} ion homeostasis [162, 163]. This provides a direct link between mitochondrial damage and intracellular Ca^{2+} overload. Mitochondria serve as primary site for the generation of the oxidative and nitrosative stress. The proximity of mtDNA to the sites of ROS and RNS generation and lack of protective histones makes it highly vulnerable for ROS-induced damage [164]. Literature evidences suggest that ROS- and RNS-mediated cellular damage activates glial cells to produce NO and pro-inflammatory cytokines with subsequent formation of RNS [147, 155]. Activated glial cells also release H_2O_2 , which is converted to hypochlorous acid (HOCl), a highly toxic pro-oxidant [165]. Thus, a vicious cycle of Ca^{2+} overload, oxidative stress and inflammation mediates the progression of neurodegenerative diseases. This makes it challenging to distinguish the sequence of the events.

3. NUTRACEUTICALS: BRIEF OVERVIEW

Although, our understanding about the mechanisms involved in the neuronal damage has increased in recent years, we are yet to identify natural compound-based therapeutic agents that positively improve the overall neuronal health. One of the major disadvantages of the currently available therapeutic agents against the neuronal damage is that chronic use of these agents results in the multiple adverse effects. Therefore, protective strategies including nutraceuticals-based and other cost-effective therapies are warranted. The term "Nutraceutical" was coined by Dr. Stephen DeFelice, the then Chairman of the Foundation for Innovation Medicine, in 1989 [166]. Nutraceuticals are food-based products that not only supplement the diet but also help in the prevention and/or treatment of a wide variety of diseases and disorders [167]. About 2000 years ago, Hippocrates correctly stated that "Let

food be your medicine and medicine be your food" [168]. According to the epidemiological studies carried out in the 1980s by Professors Doll and Peto for the World Health Organization, it has been postulated that appropriate nutrition could prevent approximately 35% of cancer deaths, and as many as 90% of certain cancers could be avoided by dietary supplementation [169]. As stated above, several mechanistic studies found that mitotoxicity, intracellular Ca^{2+} overload, oxidative stress and inflammation are the primary reasons for the number of neurodegenerative disorders including AD, PD, HD, MS and ALS [55, 96, 111, 142]. Nutraceuticals by virtue of their better tolerance and ability to act on multiple cellular pathways appear to be favorable candidates for the long-term consumption as required for the management of chronic ailments such as neurodegenerative disorders. Currently nutraceuticals have received considerable global interest because of their presumed safety and potential nutritional and therapeutic effects.

Some nutraceuticals such as resveratrol, α -lipoic acid, coenzyme Q10 (ubiquinone), β -carotene, lycopene, astaxanthin and curcumin, to name a few, are potent antioxidants and have showed therapeutic effects against several diseases by neutralizing oxidative stress, boosting the endogenous antioxidant levels and stabilizing mitochondrial functions [37-39, 170-173]. Several studies have suggested beneficial effects of a wide variety of nutraceuticals against various neurodegenerative disorders; however, the present review focuses on nutraceuticals for which mechanistic evidences for neuroprotection are available (Table 1).

In general, the neuroprotective effects of nutraceuticals can be attributed, but not limited, to their ability to scavenge free radicals, chelate transition metals, improve antioxidant reserve and anti-inflammatory effects. At the molecular level, nutraceuticals regulate several signaling pathways which are known to play role in the cell survival and stress response such as Nrf2/ARE, mitogen-activated protein kinase (MAPK) [174], protein kinase C (PKC) [175], Janus kinase-Signal Transducer and Activator of Transcription (JAK-STAT) [176, 177], MEK/ERK/CREB, PI3K/AKT [178, 179] and insulin-signaling pathways [180, 181]. As nutraceuticals act on multiple molecular pathways, they are anticipated to improve overall neuronal health. Nonetheless, we have categorized the available evidences for some nutraceuticals-mediated neuroprotection (Table 1) based on the primary effect as concluded from the respective study. Major pathways involved in neuronal damage and nutraceuticals-mediated neuroprotection are depicted in Fig. 1.

3.1. Nutraceuticals Targeting Mitochondrial Dysfunction

Neurons are vulnerable to mitochondrial damage [182, 183]. Several nutritional supplements have been tested for their ability to preserve mitochondrial functions and thereby offer protection against the neurodegenerative diseases. Curcumin, derived from the turmeric, ameliorated 6-hydroxydopamine-induced neurotoxicity in MES23.5 cells by partially restoring the $\Delta\Psi_m$, increasing the level of Cu-Zn SOD and suppressing an increase in intracellular ROS and translocation of NF- κ B, a transcription-factor involved in inflammation [55]. α -lipoic acid, another potent mitochondrial stabilizer, protected neurons *in vitro* and *in vivo*

against the chemotherapy, hypoxia, A β and other toxicants-induced neurotoxicity by improving the mitochondrial functions and physiology [184-189]. Several clinical trials have been initiated or completed for α -lipoic acid in patients with diabetic neuropathy [190] and peripheral neuropathy associated with the cancer chemotherapy [191]. Astaxanthin protected mitochondria of the cultured nerve cells and boosted energy production, without increasing the production of ROS [192, 193]. Coenzyme Q10 is another powerful mitochondrial antioxidant that protects neuronal cells by preserving the functions of mitochondria in stroke, epilepsy, striatal excitotoxic lesions produced by the mitochondrial toxin (Complex II inhibitor) and several other neurodegenerative disorders [67, 194-196]. Several clinical trials with Coenzyme Q10 suggested its beneficial effects in neurodegenerative disorders [197, 198]. These evidences strongly substantiate the ability of nutraceuticals to fight against the onset and progression of neurodegenerative disorders by primarily preserving the mitochondrial functions.

3.2. Nutraceuticals Targeting Calcium Overload

The neuroprotective effect of blue berry-extract against A β was mediated by its ability to antagonize increase in intracellular Ca^{2+} and aggregated A β -induced ATP leakage [207, 208]. Despite being a potentially effective therapeutic strategy [227], only few studies are available that examine the ability of nutraceuticals to antagonize intracellular Ca^{2+} overload. With the help of advanced *in-silico* tools, further mechanistic studies are required to identify some potential nutraceuticals that can prevent the pathological rise in intracellular $[\text{Ca}^{2+}]$.

3.3. Nutraceuticals Targeting Oxidative Stress

A large number of nutraceuticals exert both direct and indirect antioxidant effects by scavenging ROS and inducing the expression of cytoprotective proteins in a Nrf2-dependent manner, respectively [37-39]. Nrf2-antioxidant response element (ARE) signaling pathway has been identified as a promising therapeutic target for neurodegenerative diseases [228].

Pre-treatment with L-sulforaphane, an isothiocyanate compound found in broccoli and other cruciferous vegetables, markedly inhibited dopamine quinone-induced neuronal death by decreasing accumulation of ROS, attenuating membrane damage and preventing DNA fragmentation in dopaminergic cell lines (Cath. and SK-N-BE (2)C) and in mesencephalic dopaminergic neurons. Also, L-sulforaphane and tert-butylhydroquinone protected neurons and astrocytes against H_2O_2 -induced oxidative stress in a mixed neuron-astrocyte culture system through stimulation of the Nrf2-antioxidant response element (ARE) transcriptional pathway. Activation of Nrf2 pathway induces transcription of various antioxidant genes such as γ -glutamylcysteine ligase (GCL), the rate limiting enzyme in the synthesis of glutathione (GSH), MnSOD, hemoxygenase and NAD(P) H:quininereductase [205, 229]. Blue berry diet is also known to exert neuroprotection through alteration in ROS signaling through CREB and MAP-kinase signaling pathways [206]. Possible mechanisms for the neuroprotective effect of resveratrol are related to its antioxidant properties and its

Table 1. Summary of the molecular neuronal effects of emerging nutraceuticals.

S. N.	Nutraceuticals	Targeted Mechanism(s)	Implication	Refs.
1.	Curcumin	<ul style="list-style-type: none"> Restored the mitochondrial membrane potential, Suppressed increase in intracellular ROS Inhibited pro-inflammatory signaling through NF-κB or toll-like receptors Inhibited Aβ-induced MAP kinase activation and the phosphorylation of ERK-1/2 Inhibited phosphorylation of JNK1/2 and c-Jun Decreased malondialdehyde levels, cytochrome c and cleaved caspase-3 expression and increasing mitochondrial Bcl-2 expression 	<ul style="list-style-type: none"> Ameliorated 6-hydroxydopamine-induced neurotoxicity in MES23.5 cells Inhibited pro-apoptotic signals in mouse models of encephalitis Prevented Aβ-induced cell death in a human neuroblastoma cell line Stabilized the blood brain barrier in MS Protected C57BL/6 N mice and SH-SY5Y cells against dopaminergic neurotoxicity induced by MPTP or 1-methyl-4-phenylpyridinium ion- (MPP (+)) Protected rats against stroke 	[17, 71, 199-204]
2.	α -lipoic acid	<ul style="list-style-type: none"> Improved mitochondrial functions and physiology 	Protected neurons <i>in vitro</i> and <i>in vivo</i> against the chemotherapy, hypoxia, A β , diabetic neuropathy and peripheral neuropathy associated with the cancer chemotherapy	[184-191]
3.	Astaxanthin	Boosted energy production by protecting mitochondria	Protected cultured nerve cells	[192, 193]
4.	Coenzyme Q10 (ubiquinone)	Preserved mitochondrial functions	Protection against stroke, epilepsy, striatal excitotoxic lesions produced by the mitochondrial toxin (Complex II inhibitor), malonate and several other neurodegenerative disorders	[56, 131, 194-196, 198]
5.	L-sulforaphane (isothiocyanate compound)	Decreased ROS and inhibited pro-inflammatory signaling through NF- κ B or toll-like receptors	<ul style="list-style-type: none"> Inhibited dopamine quinone-induced neuronal death Stabilized the blood brain barrier in MS 	[202, 205]
6.	Tert-butyl hydroquinone	Stimulated Nrf2-ARE transcriptional pathway	Protected neurons and astrocytes against H ₂ O ₂ -induced oxidative stress	[205]
7.	Blue berry	<ul style="list-style-type: none"> Altered ROS signaling through CREB and MAP-kinase signaling pathways Modulated inflammatory genes expression Antagonized the increase in intracellular Ca²⁺ activity 		[206-209]
8.	Resveratrol	<ul style="list-style-type: none"> Antioxidant properties-mediated modulations of Aβ processing and up-regulation of the longevity-linked gene, sirtuin1 Increased 5-HT activity 	Antidepressant properties	[210, 211]
9.	Carnosic acid	Scavenged ROS	Neuroprotective action both in <i>in vitro</i> models of neuronal death and in <i>in vivo</i> models of neurodegeneration	[201, 212, 213]
10.	Rosmarinic acid		Protected neuroblastoma cells from H ₂ O ₂ -induced oxidative stress	
11.	Aged garlic extract	Suppressed ROS generation and attenuated caspase-3 activation, DNA fragmentation and PARP cleavage	Protected PC12 cells against A β peptide-induced apoptosis	[214]
12.	Eugenol	Decreased 6-hydroxydopamine-induced lipid peroxidation and increased GSH level	Prevented 6-hydroxydopamine-induced reduction in the dopamine level in the mouse striatum	[215]
13.	Anthocyanins	Negatively regulated pro-oxidants and pro-inflammatory cytokines signaling pathways	Neuroprotection	[209, 216, 217]

Table 1. contd....

S. N.	Nutraceuticals	Targeted mechanism(s)	Implication	Refs.
14.	The green tea flavonoid, epigallocatechin-3-gallate	Inhibited pro-inflammatory signaling through NF-κB or toll-like receptors	Stabilized the blood brain barrier in MS	[202]
15.	Mustard oil glycoside			
16.	Retinoic acid	Decreased inflammation through balancing T-lymphocyte populations in peripheral blood	Improve plasticity, regeneration, cognition and behaviour in MS patients	[218]
17.	Vitamin D	Anti-inflammatory activity	Protective in patients with MS, PD and AD	[219]
18.	Vitamin E	Decreased pro-inflammatory cytokines such as IL-1β and TNF-α	Decreased neuroinflammation and neuronal degeneration in the brain of rat with kainic acid-induced status epilepticus	[220]
19.	Omega-3 polyunsaturated fatty acid	Reduced inflammation	Improved neurologic recovery and attenuated white matter injury after experimental traumatic brain injury	[221]
20.	Apigenin	<ul style="list-style-type: none"> • Stabilized mitochondrial membrane potential • Modulated GABAergic and glutamatergic transmission 	Protected neurons against copper-induced Aβ-mediated toxicity	[222, 223]
21.	Soy isoflavones	Modulated brain cholinergic system	Ameliorated age-related neuronal loss and cognition decline in male rats	[224]
22.	Isoflavones	Mimicked the effects of estrogen through estrogen receptor β in the brain	Improved cognitive function	[225]
23.	Emodin (3-methyl-1,6,8-trihydroxyanthraquinone), an anthraquinone derivative	Attenuated EGF receptor signaling	Protected rats against psychosis and ameliorating behavioral deficits	[226]

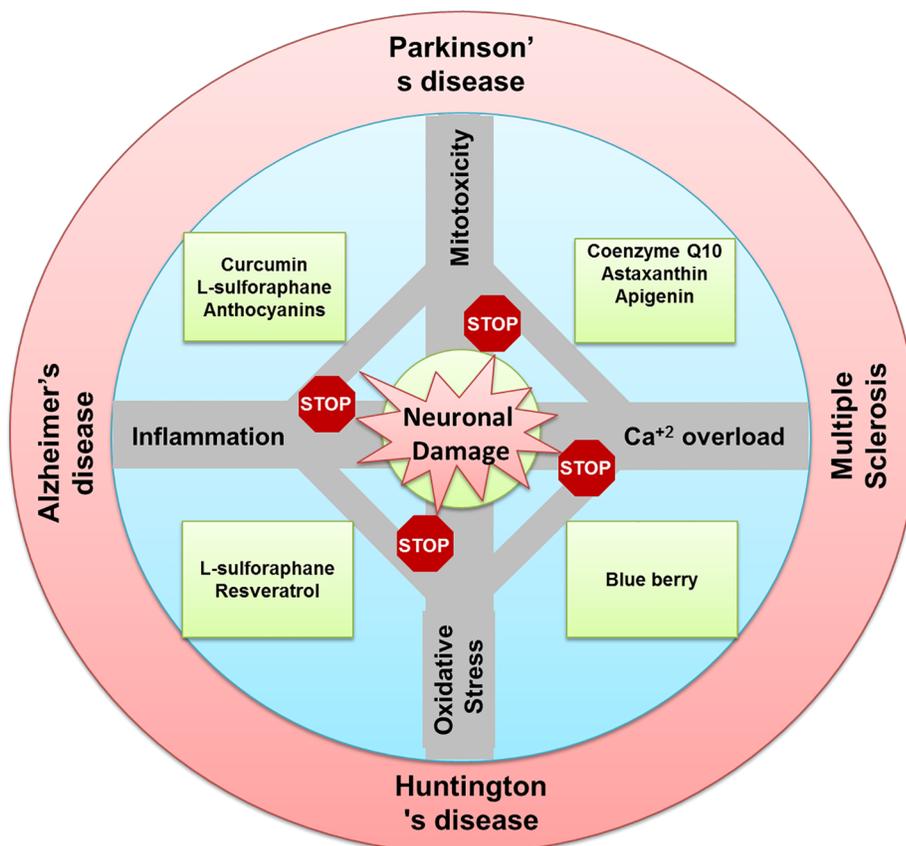


Fig. (1). A diagrammatic representation of the major pathways involved in neuronal damage and nutraceuticals-mediated neuroprotection.

capacity to modulate A β processing and up-regulation of the longevity-linked gene, sirtuin1 [211]. Curcumin decreased ROS and inhibited pro-apoptotic signals in mouse models of encephalitis [199, 201]. Carnosic acid and rosmarinic acid showed a neuroprotective action both *in vitro* and *in vivo* by scavenging ROS [201, 213]. Aged garlic extract protected PC12 cells against A β peptide-induced apoptosis by suppressing the generation of ROS and attenuating caspase-3 activation, DNA fragmentation and PARP cleavage [214]. Eugenol, a nutraceutical obtained from cloves, prevented 6-hydroxydopamine-induced reduction in the dopamine level in the mouse striatum by decreasing 6-hydroxydopamine-induced lipid peroxidation and increasing the GSH level [215]. Epidemiological studies have demonstrated that consuming foods rich in antioxidant vitamin supplements such as vitamin C and E are associated with a lower risk of developing PD [230]. However, at present, we are not aware of any conclusive clinical trials for these antioxidants against neurodegenerative disorders.

3.4. Nutraceuticals Targeting Inflammation

The neuroprotective effect of anthocyanins is known to be mediated through phospholipase A2 inhibition [216], which is adversely involved in a complex network of signaling pathways linking pro-oxidants and pro-inflammatory cytokines to the release of arachidonic acid and eicosanoid synthesis [217]. Blue berry diet showed neuroprotection through the modulation of genes expression involved in inflammation [209]. Curcumin inhibited the activation of NF- κ B and prevented A β -induced cell death in a human neuroblastoma cell line, suggesting its possible role in the treatment for AD [11]. The green tea flavonoid, epigallocatechin-3-gallate, curcumin and the mustard oil glycoside are known to inhibit the pro-inflammatory signaling through NF- κ B or toll-like receptors and stabilize the blood brain barrier in MS [202]. Retinoic acid, a metabolite of vitamin A, is known to increase tolerance and decrease inflammation, through balancing T-lymphocyte populations in peripheral blood, and thereby may improve plasticity, regeneration, cognition and behavior in patients with MS [218]. Vitamin D has been shown to alleviate various inflammatory markers in patients with MS, PD and AD [219]. Vitamin E supplementation reduced neuroinflammation and neuronal degeneration in the brain of rat with kainic acid-induced status epilepticus by decreasing the levels of astrocytic and microglial antigens (GFAP and MHC II, respectively) and pro-inflammatory cytokines such as IL-1 β and TNF- α [220]. Omega-3 polyunsaturated fatty acid supplementation improved neurologic recovery and attenuated white matter injury after experimental traumatic brain injury by eliciting multifaceted protection against behavioral dysfunction, hippocampal neuronal loss, inflammation, and loss of myelination and impulse conductivity [221].

3.5. Nutraceuticals as Neurotransmitter Modulators

Apigenin, a flavonoid, protected neurons against copper-induced A β -mediated toxicity by relieving mitochondrial $\Delta\Psi_m$ dissipation [223] and modulating GABAergic and glutamatergic transmission in the cultured cortical neurons [222]. Moreover, soy isoflavones are known to influence the

brain cholinergic system and ameliorate age-related neuronal loss and cognition decline in male rats [224]. Resveratrol, a non-flavonoid polyphenol, possesses antidepressant properties, which could be attributed to its ability to increase 5-HT level [210]. Thus, a few compounds have been identified to possess neuromodulatory activities that can be leveraged for the treatment of neurodegenerative disorders.

4. CONCLUSIONS

Mitotoxicity, calcium overload, oxidative stress and inflammation are the common pathways that are involved, often together, in the neuronal damage. Activation of these pathways, *viz.* mitochondrial dysfunction, intracellular Ca²⁺ overload, oxidative stress and inflammation, in neuronal damage is widely acknowledged, and it must be coupled with a change in the expression and function of hundreds to thousands of genes. We believe that the experimental studies utilizing advanced microarray-based investigations to identify the ability of nutraceuticals to antagonize such changes are the need of hour. At present limited information is available in this direction and further research is needed.

CONFLICT OF INTEREST

The authors confirm that this article content has no conflict of interest.

ACKNOWLEDGEMENTS

Declared none.

REFERENCES

- [1] Fu, X.Y.; Yang, M.F.; Cao, M.Z.; Li, D.W.; Yang, X.Y.; Sun, J.Y.; Zhang, Z.Y.; Mao, L.L.; Zhang, S.; Wang, F.Z.; Zhang, F.; Fan, C.D.; Sun, B.L. Strategy to Suppress Oxidative Damage-Induced Neurotoxicity in PC12 Cells by Curcumin: the Role of ROS-Mediated DNA Damage and the MAPK and AKT Pathways. *Mol. Neurobiol.*, **2014**. [PMID: 25432891]
- [2] Keber, U.; Klietz, M.; Carlsson, T.; Oertel, W.H.; Weihe, E.; Schäfer, M.K.; Höglinger, G.U.; Depboylu, C. Striatal tyrosine hydroxylase-positive neurons are associated with L-DOPA-induced dyskinesia in hemiparkinsonian mice. *Neuroscience*, **2015**, *298*, 302-317. [PMID: 25892702]
- [3] Kumar, A.; Leinisch, F.; Kadiiska, M.B.; Corbett, J.; Mason, R.P. Formation and Implications of Alpha-Synuclein Radical in Maneb- and Paraquat-Induced Models of Parkinson's Disease. *Mol. Neurobiol.*, **2015**. [PMID: 25952542]
- [4] Mattson, M.P.; Magnus, T. Ageing and neuronal vulnerability. *Nat. Rev. Neurosci.*, **2006**, *7*(4), 278-294. [PMID: 16552414]
- [5] Zindo, F.T.; Joubert, J.; Malan, S.F. Propargylamine as functional moiety in the design of multifunctional drugs for neurodegenerative disorders: MAO inhibition and beyond. *Future Med. Chem.*, **2015**, *7*(5), 609-629. [PMID: 25921401]
- [6] Amor, S.; Puentes, F.; Baker, D.; van der Valk, P. Inflammation in neurodegenerative diseases. *Immunology*, **2010**, *129*(2), 154-169. [PMID: 20561356]
- [7] Cunningham, C. Microglia and neurodegeneration: the role of systemic inflammation. *Glia*, **2013**, *61*(1), 71-90. [PMID: 22674585]
- [8] Federico, A.; Cardaioli, E.; Da Pozzo, P.; Formichi, P.; Gallus, G.N.; Radi, E. Mitochondria, oxidative stress and neurodegeneration. *J. Neurol. Sci.*, **2012**, *322*(1-2), 254-262. [PMID: 22669122]
- [9] Glass, C.K.; Saijo, K.; Winner, B.; Marchetto, M.C.; Gage, F.H. Mechanisms underlying inflammation in neurodegeneration. *Cell*, **2010**, *140*(6), 918-934. [PMID: 20303880]
- [10] Halliwell, B. Oxidative stress and neurodegeneration: where are we now? *J. Neurochem.*, **2006**, *97*(6), 1634-1658. [PMID: 16805774]

- [11] Lin, M.T.; Beal, M.F. Mitochondrial dysfunction and oxidative stress in neurodegenerative diseases. *Nature*, **2006**, *443*(7113), 787-795. [PMID: 17051205]
- [12] Moreira, P.I.; Zhu, X.; Wang, X.; Lee, H.G.; Nunomura, A.; Petersen, R.B.; Perry, G.; Smith, M.A. Mitochondria: a therapeutic target in neurodegeneration. *Biochim. Biophys. Acta*, **2010**, *1802*(1), 212-220. [PMID: 19853657]
- [13] Nguyen, H.T.; Sawmiller, D.R.; Markov, O.; Chen, M. Elevated $[Ca^{2+}]_i$ levels occur with decreased calpain activity in aged fibroblasts and their reversal by energy-rich compounds: new paradigm for Alzheimer's disease prevention. *J. Alzheimers Dis.*, **2013**, *37*(4), 835-848. [PMID: 24122005]
- [14] Uttara, B.; Singh, A.V.; Zamboni, P.; Mahajan, R.T. Oxidative stress and neurodegenerative diseases: a review of upstream and downstream antioxidant therapeutic options. *Curr. Neuropharmacol.*, **2009**, *7*(1), 65-74. [PMID: 19721819]
- [15] Cali, T.; Ottolini, D.; Brini, M. Mitochondrial Ca^{2+} and neurodegeneration. *Cell Calcium*, **2012**, *52*(1), 73-85. [PMID: 22608276]
- [16] Celsi, F.; Pizzo, P.; Brini, M.; Leo, S.; Fotino, C.; Pinton, P.; Rizzuto, R. Mitochondria, calcium and cell death: a deadly triad in neurodegeneration. *Biochim. Biophys. Acta*, **2009**, *1787*(5), 335-344. [PMID: 19268425]
- [17] Verkhratsky, A.; Toescu, E.C. Endoplasmic reticulum Ca^{2+} homeostasis and neuronal death. *J. Cell. Mol. Med.*, **2003**, *7*(4), 351-361. [PMID: 14754504]
- [18] Liu, Y.; Fiskum, G.; Schubert, D. Generation of reactive oxygen species by the mitochondrial electron transport chain. *J. Neurochem.*, **2002**, *80*(5), 780-787. [PMID: 11948241]
- [19] Gilgun-Sherki, Y.; Melamed, E.; Offen, D. Anti-inflammatory drugs in the treatment of neurodegenerative diseases: current state. *Curr. Pharm. Des.*, **2006**, *12*(27), 3509-3519. [PMID: 17017944]
- [20] Jung, Y.W.; Hysolli, E.; Kim, K.Y.; Tanaka, Y.; Park, I.H. Human induced pluripotent stem cells and neurodegenerative disease: prospects for novel therapies. *Curr. Opin. Neurol.*, **2012**, *25*(2), 125-130. [PMID: 22357218]
- [21] Young, A.B. Four decades of neurodegenerative disease research: how far we have come! *J. Neurosci.*, **2009**, *29*(41), 12722-12728. [PMID: 19828782]
- [22] Connolly, B.S.; Lang, A.E. Pharmacological treatment of Parkinson disease: a review. *JAMA*, **2014**, *311*(16), 1670-1683. [PMID: 24756517]
- [23] Dantuma, E.; Merchant, S.; Sugaya, K. Stem cells for the treatment of neurodegenerative diseases. *Stem Cell Res. Ther.*, **2010**, *1*(5), 37. [PMID: 21144012]
- [24] Dunkel, P.; Chai, C.L.; Sperlágh, B.; Huleatt, P.B.; Mátyus, P. Clinical utility of neuroprotective agents in neurodegenerative diseases: current status of drug development for Alzheimer's, Parkinson's and Huntington's diseases, and amyotrophic lateral sclerosis. *Expert Opin. Investig. Drugs*, **2012**, *21*(9), 1267-1308. [PMID: 22741814]
- [25] Dye, R.V.; Miller, K.J.; Singer, E.J.; Levine, A.J. Hormone replacement therapy and risk for neurodegenerative diseases. *Int. J. Alzheimers Dis.*, **2012**, *2012*, 258454.
- [26] Moraes, W.A. Current Pharmacological and Non-Pharmacological Therapies for Neurodegenerative Diseases. **2015**, 23-48.
- [27] Moreno, J.A.; Halliday, M.; Molloy, C.; Radford, H.; Verity, N.; Axten, J.M.; Ortori, C.A.; Willis, A.E.; Fischer, P.M.; Barrett, D.A.; Mallucci, G.R. Oral treatment targeting the unfolded protein response prevents neurodegeneration and clinical disease in prion-infected mice. *Sci. Transl. Med.*, **2013**, *5*(206), 206ra138. [PMID: 24107777]
- [28] Weissmiller, A.M.; Wu, C. Current advances in using neurotrophic factors to treat neurodegenerative disorders. *Transl. Neurodegener.*, **2012**, *1*(1), 14. [PMID: 23210531]
- [29] Morrish, P. Prescribing in Parkinson's disease: a story of hope and adverse events. *Pract. Neurol.*, **2012**, *12*(5), 335-340. [PMID: 22976066]
- [30] Roch-Torreilles, I.; Camu, W.; Hillaire-Buys, D. [Adverse effects of riluzole (Rilutek) in the treatment of amyotrophic lateral sclerosis]. *Therapie*, **2000**, *55*(2), 303-312. [Adverse effects of riluzole (Rilutek) in the treatment of amyotrophic lateral sclerosis]. [PMID: 10967703]
- [31] Abeliovich, A.; Doege, C.A. Reprogramming therapeutics: iPS cell prospects for neurodegenerative disease. *Neuron*, **2009**, *61*(3), 337-339. [PMID: 19217371]
- [32] Lunn, J.S.; Sakowski, S.A.; Hur, J.; Feldman, E.L. Stem cell technology for neurodegenerative diseases. *Ann. Neurol.*, **2011**, *70*(3), 353-361. [PMID: 21905078]
- [33] Whalley, K. Neurodegenerative disease: Discouraging transplant results. *Nat. Rev. Neurosci.*, **2009**, *10*(9), 624-624.
- [34] Essa, M.M.; Subash, S.; Akbar, M.; Al-Adawi, S.; Guillemin, G.J. Long-term dietary supplementation of pomegranates, figs and dates alleviate neuroinflammation in a transgenic mouse model of Alzheimer's disease. *PLoS One*, **2015**, *10*(3), e0120964. [PMID: 25807081]
- [35] Pugh, N.D.; Edwall, D.; Lindmark, L.; Kousoulas, K.G.; Iyer, A.V.; Haron, M.H.; Pasco, D.S. Oral administration of a Spirulina extract enriched for Braun-type lipoproteins protects mice against influenza A (H1N1) virus infection. *Phytomedicine*, **2015**, *22*(2), 271-276. [PMID: 25765832]
- [36] Sharma, S.S.; Kumar, A.; Arora, M.; Kaundal, R.K. Neuroprotective potential of combination of resveratrol and 4-amino 1,8 naphthalimide in experimental diabetic neuropathy: focus on functional, sensorimotor and biochemical changes. *Free Radic. Res.*, **2009**, *43*(4), 400-408. [PMID: 19291593]
- [37] Tripathi, D.N.; Jena, G.B. Astaxanthin intervention ameliorates cyclophosphamide-induced oxidative stress, DNA damage and early hepatocarcinogenesis in rat: role of Nrf2, p53, p38 and phase-II enzymes. *Mutat. Res.*, **2010**, *696*(1), 69-80. [PMID: 20038455]
- [38] Trivedi, P.P.; Jena, G.B. Role of α -lipoic acid in dextran sulfate sodium-induced ulcerative colitis in mice: studies on inflammation, oxidative stress, DNA damage and fibrosis. *Food Chem. Toxicol.*, **2013**, *59*, 339-355. [PMID: 23793040]
- [39] Trivedi, P.P.; Jena, G.B. Mechanistic insight into beta-carotene-mediated protection against ulcerative colitis-associated local and systemic damage in mice. *Eur. J. Nutr.*, **2014**. [PMID: 25074825]
- [40] Atlante, A.; Calissano, P.; Bobba, A.; Giannattasio, S.; Marra, E.; Passarella, S. Glutamate neurotoxicity, oxidative stress and mitochondria. *FEBS Lett.*, **2001**, *497*(1), 1-5. [PMID: 11376653]
- [41] Chen, H.; Chan, D.C. Mitochondrial dynamics--fusion, fission, movement, and mitophagy--in neurodegenerative diseases. *Hum. Mol. Genet.*, **2009**, *18*(R2), R169-R176. [PMID: 19808793]
- [42] Galluzzi, L.; Blomgren, K.; Kroemer, G. Mitochondrial membrane permeabilization in neuronal injury. *Nat. Rev. Neurosci.*, **2009**, *10*(7), 481-494. [PMID: 19543220]
- [43] Mancuso, M.; Coppede, F.; Migliore, L.; Siciliano, G.; Murri, L. Mitochondrial dysfunction, oxidative stress and neurodegeneration. *J. Alzheimers Dis.*, **2006**, *10*(1), 59-73. [PMID: 16988483]
- [44] Rintoul, G.L.; Reynolds, I.J. Mitochondrial trafficking and morphology in neuronal injury. *Biochim. Biophys. Acta*, **2010**, *1802*(1), 143-150. [PMID: 19747973]
- [45] Schinder, A.F.; Olson, E.C.; Spitzer, N.C.; Montal, M. Mitochondrial dysfunction is a primary event in glutamate neurotoxicity. *J. Neurosci.*, **1996**, *16*(19), 6125-6133. [PMID: 8815895]
- [46] Su, B.; Wang, X.; Zheng, L.; Perry, G.; Smith, M.A.; Zhu, X. Abnormal mitochondrial dynamics and neurodegenerative diseases. *Biochim. Biophys. Acta*, **2010**, *1802*(1), 135-142. [PMID: 19799998]
- [47] White, R.J.; Reynolds, I.J. Mitochondria and Na^{+}/Ca^{2+} exchange buffer glutamate-induced calcium loads in cultured cortical neurons. *J. Neurosci.*, **1995**, *15*(2), 1318-1328. [PMID: 7869100]
- [48] Wieloch, T. Mitochondrial involvement in acute neurodegeneration. *IUBMB Life*, **2001**, *52*(3-5), 247-254. [PMID: 11798039]
- [49] Beal, M.F. Mitochondria take center stage in aging and neurodegeneration. *Ann. Neurol.*, **2005**, *58*(4), 495-505. [PMID: 16178023]
- [50] Nicklas, W.J.; Vyas, I.; Heikkila, R.E. Inhibition of NADH-linked oxidation in brain mitochondria by 1-methyl-4-phenyl-pyridine, a metabolite of the neurotoxin, 1-methyl-4-phenyl-1,2,5,6-tetrahydropyridine. *Life Sci.*, **1985**, *36*(26), 2503-2508. [PMID: 2861548]
- [51] Ramsay, R.R.; Salach, J.I.; Dadgar, J.; Singer, T.P. Inhibition of mitochondrial NADH dehydrogenase by pyridine derivatives and its possible relation to experimental and idiopathic parkinsonism. *Biochem. Biophys. Res. Commun.*, **1986**, *135*(1), 269-275. [PMID: 3485428]
- [52] Zheng, H.; Xiao, W.H.; Bennett, G.J. Functional deficits in peripheral nerve mitochondria in rats with paclitaxel- and

- oxaliplatin-evoked painful peripheral neuropathy. *Exp. Neurol.*, **2011**, 232(2), 154-161. [PMID: 21907196]
- [53] Zheng, H.; Xiao, W.H.; Bennett, G.J. Mitotoxicity and bortezomib-induced chronic painful peripheral neuropathy. *Exp. Neurol.*, **2012**, 238(2), 225-234. [PMID: 22947198]
- [54] Podratz, J.L.; Knight, A.M.; Ta, L.E.; Staff, N.P.; Gass, J.M.; Genel, K.; Schlattau, A.; Lathroum, L.; Windebank, A.J. Cisplatin induced mitochondrial DNA damage in dorsal root ganglion neurons. *Neurobiol. Dis.*, **2011**, 41(3), 661-668. [PMID: 21145397]
- [55] Aliev, G.; Seyidova, D.; Neal, M.L.; Shi, J.; Lamb, B.T.; Siedlak, S.L.; Vinters, H.V.; Head, E.; Perry, G.; Lamanna, J.C.; Friedland, R.P.; Cotman, C.W. Atherosclerotic lesions and mitochondria DNA deletions in brain microvessels as a central target for the development of human AD and AD-like pathology in aged transgenic mice. *Ann. N. Y. Acad. Sci.*, **2002**, 977, 45-64. [PMID: 12480733]
- [56] Beal, M.F. Neurochemistry and toxin models in Huntington's disease. *Curr. Opin. Neurol.*, **1994**, 7(6), 542-547. [PMID: 7866587]
- [57] Beal, M.F. Mitochondrial dysfunction and oxidative damage in Alzheimer's and Parkinson's diseases and coenzyme Q10 as a potential treatment. *J. Bioenerg. Biomembr.*, **2004**, 36(4), 381-386. [PMID: 15377876]
- [58] Su, K.; Bourdette, D.; Forte, M. Mitochondrial dysfunction and neurodegeneration in multiple sclerosis. *Front. Physiol.*, **2013**, 4, 169. [PMID: 23898299]
- [59] Wong, P.C.; Pardo, C.A.; Borchelt, D.R.; Lee, M.K.; Copeland, N.G.; Jenkins, N.A.; Sisodia, S.S.; Cleveland, D.W.; Price, D.L. An adverse property of a familial ALS-linked SOD1 mutation causes motor neuron disease characterized by vacuolar degeneration of mitochondria. *Neuron*, **1995**, 14(6), 1105-1116. [PMID: 7605627]
- [60] Dawson, T.M.; Dawson, V.L. Molecular pathways of neurodegeneration in Parkinson's disease. *Science*, **2003**, 302(5646), 819-822. [PMID: 14593166]
- [61] Schapira, A.H. Causes of neuronal death in Parkinson's disease. *Adv. Neurol.*, **2001**, 86, 155-162. [PMID: 11553973]
- [62] Sullivan, P.G.; Brown, M.R. Mitochondrial aging and dysfunction in Alzheimer's disease. *Prog. Neuropsychopharmacol. Biol. Psychiatry*, **2005**, 29(3), 407-410. [PMID: 15795049]
- [63] Brown, R.H., Jr. Amyotrophic lateral sclerosis: recent insights from genetics and transgenic mice. *Cell*, **1995**, 80(5), 687-692. [PMID: 7889564]
- [64] Hensley, K.; Mhatre, M.; Mou, S.; Pye, Q.N.; Stewart, C.; West, M.; Williamson, K.S. On the relation of oxidative stress to neuroinflammation: lessons learned from the G93A-SOD1 mouse model of amyotrophic lateral sclerosis. *Antioxid. Redox Signal.*, **2006**, 8(11-12), 2075-2087. [PMID: 17034351]
- [65] Di Filippo, M.; Tozzi, A.; Tantucci, M.; Arcangeli, S.; Chiasserini, D.; Ghiglieri, V.; de Iure, A.; Calabresi, P. Interferon- β 1 protects neurons against mitochondrial toxicity via modulation of STAT1 signaling: electrophysiological evidence. *Neurobiol. Dis.*, **2014**, 62, 387-393. [PMID: 24135008]
- [66] Qi, X.; Lewin, A.S.; Sun, L.; Hauswirth, W.W.; Guy, J. Mitochondrial protein nitration primes neurodegeneration in experimental autoimmune encephalomyelitis. *J. Biol. Chem.*, **2006**, 281(42), 31950-31962. [PMID: 16920708]
- [67] Beal, M.F.; Henshaw, D.R.; Jenkins, B.G.; Rosen, B.R.; Schulz, J.B. Coenzyme Q10 and nicotinamide block striatal lesions produced by the mitochondrial toxin malonate. *Ann. Neurol.*, **1994**, 36(6), 882-888. [PMID: 7998775]
- [68] Beal, M.F.; Swartz, K.J.; Hyman, B.T.; Storey, E.; Finn, S.F.; Koroshetz, W. Aminooxyacetic acid results in excitotoxic lesions by a novel indirect mechanism. *J. Neurochem.*, **1991**, 57(3), 1068-1073. [PMID: 1830613]
- [69] Greene, J.G.; Porter, R.H.; Eller, R.V.; Greenamyre, J.T. Inhibition of succinate dehydrogenase by malonic acid produces an "excitotoxic" lesion in rat striatum. *J. Neurochem.*, **1993**, 61(3), 1151-1154. [PMID: 8360680]
- [70] Ludolph, A.C.; He, F.; Spencer, P.S.; Hammerstad, J.; Sabri, M. 3-Nitropropionic acid-exogenous animal neurotoxin and possible human striatal toxin. *Can. J. Neurol. Sci.*, **1991**, 18(4), 492-498. [PMID: 1782616]
- [71] Tang, T.S.; Slow, E.; Lupu, V.; Stavrovskaya, I.G.; Sugimori, M.; Llinás, R.; Kristal, B.S.; Hayden, M.R.; Bezprozvanny, I. Disturbed Ca²⁺ signaling and apoptosis of medium spiny neurons in Huntington's disease. *Proc. Natl. Acad. Sci. USA*, **2005**, 102(7), 2602-2607. [PMID: 15695335]
- [72] Gu, M.; Gash, M.T.; Mann, V.M.; Javoy-Agid, F.; Cooper, J.M.; Schapira, A.H. Mitochondrial defect in Huntington's disease caudate nucleus. *Ann. Neurol.*, **1996**, 39(3), 385-389. [PMID: 8602759]
- [73] Reddy, P.H.; Shirendeb, U.P. Mutant huntingtin, abnormal mitochondrial dynamics, defective axonal transport of mitochondria, and selective synaptic degeneration in Huntington's disease. *Biochim. Biophys. Acta*, **2012**, 1822(2), 101-110. [PMID: 22080977]
- [74] Trushina, E.; McMurray, C.T. Oxidative stress and mitochondrial dysfunction in neurodegenerative diseases. *Neuroscience*, **2007**, 145(4), 1233-1248. [PMID: 17303344]
- [75] Schanne, F.A.; Kane, A.B.; Young, E.E.; Farber, J.L. Calcium dependence of toxic cell death: a final common pathway. *Science*, **1979**, 206(4419), 700-702. [PMID: 386513]
- [76] Schlaepfer, W.W.; Bunge, R.P. Effects of calcium ion concentration on the degeneration of amputated axons in tissue culture. *J. Cell Biol.*, **1973**, 59(2 Pt 1), 456-470. [PMID: 4805010]
- [77] Borle, A.B. Membrane transfer of calcium. *Clin. Orthop. Relat. Res.*, **1967**, 52(52), 267-291. [PMID: 6048916]
- [78] Hodgkin, A.L.; Keynes, R.D. Movements of labelled calcium in squid giant axons. *J. Physiol.*, **1957**, 138(2), 253-281. [PMID: 13526124]
- [79] Rasmussen, H. Cell communication, calcium ion, and cyclic adenosine monophosphate. *Science*, **1970**, 170(3956), 404-412. [PMID: 4318972]
- [80] Blaustein, M.P.; Lederer, W.J. Sodium/calcium exchange: its physiological implications. *Physiol. Rev.*, **1999**, 79(3), 763-854. [PMID: 10390518]
- [81] Tatsumi, H.; Katayama, Y. Regulation of the intracellular free calcium concentration in acutely dissociated neurones from rat nucleus basalis. *J. Physiol.*, **1993**, 464, 165-181. [PMID: 8229797]
- [82] Lehninger, A.L. Mitochondria and calcium ion transport. *Biochem. J.*, **1970**, 119(2), 129-138. [PMID: 4922961]
- [83] Racay, P.; Kaplán, P.; Lehotský, J. Control of Ca²⁺ homeostasis in neuronal cells. *Gen. Physiol. Biophys.*, **1996**, 15(3), 193-210. [PMID: 9076503]
- [84] Orrenius, S.; Zhivotovsky, B.; Nicotera, P. Regulation of cell death: the calcium-apoptosis link. *Nat. Rev. Mol. Cell Biol.*, **2003**, 4(7), 552-565. [PMID: 12838338]
- [85] Chao, D.; Xia, Y. Ionic storm in hypoxic/ischemic stress: can opioid receptors subside it? *Prog. Neurobiol.*, **2010**, 90(4), 439-470. [PMID: 20036308]
- [86] Simon, R.P.; Griffiths, T.; Evans, M.C.; Swan, J.H.; Meldrum, B.S. Calcium overload in selectively vulnerable neurons of the hippocampus during and after ischemia: an electron microscopy study in the rat. *J. Cereb. Blood Flow Metab.*, **1984**, 4(3), 350-361. [PMID: 6470053]
- [87] Murphy, A.N.; Fiskum, G.; Beal, M.F. Mitochondria in neurodegeneration: bioenergetic function in cell life and death. *J. Cereb. Blood Flow Metab.*, **1999**, 19(3), 231-245. [PMID: 10078875]
- [88] Penn, R.D.; Loewenstein, W.R. Uncoupling of a nerve cell membrane junction by calcium-ion removal. *Science*, **1966**, 151(3706), 88-89. [PMID: 4956065]
- [89] Tedesco, E.; Rigoni, M.; Caccin, P.; Grishin, E.; Rossetto, O.; Montecucco, C. Calcium overload in nerve terminals of cultured neurons intoxicated by alpha-latrotoxin and snake PLA2 neurotoxins. *Toxicon*, **2009**, 54(2), 138-144. [PMID: 19341756]
- [90] Manev, H.; Favaron, M.; Guidotti, A.; Costa, E. Delayed increase of Ca²⁺ influx elicited by glutamate: role in neuronal death. *Mol. Pharmacol.*, **1989**, 36(1), 106-112. [PMID: 2568579]
- [91] Marcoux, F.W.; Probert, A.W., Jr; Weber, M.L. Hypoxic neuronal injury in tissue culture is associated with delayed calcium accumulation. *Stroke*, **1990**, 21(11)(Suppl.), III71-III74. [PMID: 2146782]
- [92] Mattson, M.P. Calcium and neurodegeneration. *Aging Cell*, **2007**, 6(3), 337-350. [PMID: 17328689]
- [93] Zündorf, G.; Reiser, G. Calcium dysregulation and homeostasis of neural calcium in the molecular mechanisms of neurodegenerative diseases provide multiple targets for neuroprotection. *Antioxid. Redox Signal.*, **2011**, 14(7), 1275-1288. [PMID: 20615073]

- [94] Berridge, M.J. Calcium hypothesis of Alzheimer's disease. *Pflugers Arch.*, **2010**, *459*(3), 441-449. [PMID: 19795132]
- [95] Yamamoto, S.; Wajima, T.; Hara, Y.; Nishida, M.; Mori, Y. Transient receptor potential channels in Alzheimer's disease. *Biochim. Biophys. Acta*, **2007**, *1772*(8), 958-967. [PMID: 17490865]
- [96] Chan, C.S.; Gertler, T.S.; Surmeier, D.J. Calcium homeostasis, selective vulnerability and Parkinson's disease. *Trends Neurosci.*, **2009**, *32*(5), 249-256. [PMID: 19307031]
- [97] Kurnellas, M.P.; Donahue, K.C.; Elkabes, S. Mechanisms of neuronal damage in multiple sclerosis and its animal models: role of calcium pumps and exchangers. *Biochem. Soc. Trans.*, **2007**, *35*(Pt 5), 923-926. [PMID: 17956247]
- [98] Lim, D.; Fedrizzi, L.; Tartari, M.; Zuccato, C.; Cattaneo, E.; Brini, M.; Carafoli, E. Calcium homeostasis and mitochondrial dysfunction in striatal neurons of Huntington disease. *J. Biol. Chem.*, **2008**, *283*(9), 5780-5789. [PMID: 18156184]
- [99] Betteridge, D.J. What is oxidative stress? *Metabolism*, **2000**, *49*(2)(Suppl. 1), 3-8. [PMID: 10693912]
- [100] Harman, D. The aging process. *Proc. Natl. Acad. Sci. USA*, **1981**, *78*(11), 7124-7128. [PMID: 6947277]
- [101] Sayre, L.M.; Perry, G.; Smith, M.A. Oxidative stress and neurotoxicity. *Chem. Res. Toxicol.*, **2008**, *21*(1), 172-188. [PMID: 18052107]
- [102] Bush, A.I. Metals and neuroscience. *Curr. Opin. Chem. Biol.*, **2000**, *4*(2), 184-191. [PMID: 10742195]
- [103] Chance, B.; Sies, H.; Boveris, A. Hydroperoxide metabolism in mammalian organs. *Physiol. Rev.*, **1979**, *59*(3), 527-605. [PMID: 37532]
- [104] Floyd, R.A.; Carney, J.M. Free radical damage to protein and DNA: mechanisms involved and relevant observations on brain undergoing oxidative stress. *Ann. Neurol.*, **1992**, *32*(Suppl.), S22-S27. [PMID: 1510377]
- [105] Gilgun-Sherki, Y.; Melamed, E.; Offen, D. Oxidative stress induced-neurodegenerative diseases: the need for antioxidants that penetrate the blood brain barrier. *Neuropharmacology*, **2001**, *40*(8), 959-975. [PMID: 11406187]
- [106] Takahashi, S.; Takahashi, I.; Sato, H.; Kubota, Y.; Yoshida, S.; Muramatsu, Y. Age-related changes in the concentrations of major and trace elements in the brain of rats and mice. *Biol. Trace Elem. Res.*, **2001**, *80*(2), 145-158. [PMID: 11437180]
- [107] Zaleska, M.M.; Floyd, R.A. Regional lipid peroxidation in rat brain *in vitro*: possible role of endogenous iron. *Neurochem. Res.*, **1985**, *10*(3), 397-410. [PMID: 4000395]
- [108] Keeney, P.M.; Xie, J.; Capaldi, R.A.; Bennett, J.P., Jr Parkinson's disease brain mitochondrial complex I has oxidatively damaged subunits and is functionally impaired and misassembled. *J. Neurosci.*, **2006**, *26*(19), 5256-5264. [PMID: 16687518]
- [109] Quinton, M.S.; Yamamoto, B.K. Causes and consequences of methamphetamine and MDMA toxicity. *AAPS J.*, **2006**, *8*(2), E337-E347. [PMID: 16796384]
- [110] Yamamoto, B.K.; Bankson, M.G. Amphetamine neurotoxicity: cause and consequence of oxidative stress. *Crit. Rev. Neurobiol.*, **2005**, *17*(2), 87-117. [PMID: 16808729]
- [111] Hensley, K.; Butterfield, D.A.; Hall, N.; Cole, P.; Subramaniam, R.; Mark, R.; Mattson, M.P.; Markesbery, W.R.; Harris, M.E.; Aksenov, M. Reactive oxygen species as causal agents in the neurotoxicity of the Alzheimer's disease-associated amyloid beta peptide. *Ann. N. Y. Acad. Sci.*, **1996**, *786*, 120-134. [PMID: 8687014]
- [112] Liu, Q.; Smith, M.A.; Avilá, J.; DeBernardis, J.; Kansal, M.; Takeda, A.; Zhu, X.; Nunomura, A.; Honda, K.; Moreira, P.I.; Oliveira, C.R.; Santos, M.S.; Shimohama, S.; Aliev, G.; de la Torre, J.; Ghanbari, H.A.; Siedlak, S.L.; Harris, P.L.; Sayre, L.M.; Perry, G. Alzheimer-specific epitopes of tau represent lipid peroxidation-induced conformations. *Free Radic. Biol. Med.*, **2005**, *38*(6), 746-754. [PMID: 15721985]
- [113] Nunomura, A.; Perry, G.; Aliev, G.; Hirai, K.; Takeda, A.; Balraj, E.K.; Jones, P.K.; Ghanbari, H.; Wataya, T.; Shimohama, S.; Chiba, S.; Atwood, C.S.; Petersen, R.B.; Smith, M.A. Oxidative damage is the earliest event in Alzheimer disease. *J. Neuropathol. Exp. Neurol.*, **2001**, *60*(8), 759-767. [PMID: 11487050]
- [114] Nunomura, A.; Perry, G.; Pappolla, M.A.; Wade, R.; Hirai, K.; Chiba, S.; Smith, M.A. RNA oxidation is a prominent feature of vulnerable neurons in Alzheimer's disease. *J. Neurosci.*, **1999**, *19*(6), 1959-1964. [PMID: 10066249]
- [115] Perry, G.; Raina, A.K.; Nunomura, A.; Wataya, T.; Sayre, L.M.; Smith, M.A. How important is oxidative damage? Lessons from Alzheimer's disease. *Free Radic. Biol. Med.*, **2000**, *28*(5), 831-834. [PMID: 10754280]
- [116] Sayre, L.M.; Zelasko, D.A.; Harris, P.L.; Perry, G.; Salomon, R.G.; Smith, M.A. 4-Hydroxynonenal-derived advanced lipid peroxidation end products are increased in Alzheimer's disease. *J. Neurochem.*, **1997**, *68*(5), 2092-2097. [PMID: 9109537]
- [117] Alam, Z.I.; Jenner, A.; Daniel, S.E.; Lees, A.J.; Cairns, N.; Marsden, C.D.; Jenner, P.; Halliwell, B. Oxidative DNA damage in the parkinsonian brain: an apparent selective increase in 8-hydroxyguanine levels in substantia nigra. *J. Neurochem.*, **1997**, *69*(3), 1196-1203. [PMID: 9282943]
- [118] Kikuchi, A.; Takeda, A.; Onodera, H.; Kimpara, T.; Hisanaga, K.; Sato, N.; Nunomura, A.; Castellani, R.J.; Perry, G.; Smith, M.A.; Itoyama, Y. Systemic increase of oxidative nucleic acid damage in Parkinson's disease and multiple system atrophy. *Neurobiol. Dis.*, **2002**, *9*(2), 244-248. [PMID: 11895375]
- [119] Zhang, J.; Perry, G.; Smith, M.A.; Robertson, D.; Olson, S.J.; Graham, D.G.; Montine, T.J. Parkinson's disease is associated with oxidative damage to cytoplasmic DNA and RNA in substantia nigra neurons. *Am. J. Pathol.*, **1999**, *154*(5), 1423-1429. [PMID: 10329595]
- [120] Alam, Z.I.; Daniel, S.E.; Lees, A.J.; Marsden, D.C.; Jenner, P.; Halliwell, B. A generalised increase in protein carbonyls in the brain in Parkinson's but not incidental Lewy body disease. *J. Neurochem.*, **1997**, *69*(3), 1326-1329. [PMID: 9282961]
- [121] Floor, E.; Wetzel, M.G. Increased protein oxidation in human substantia nigra pars compacta in comparison with basal ganglia and prefrontal cortex measured with an improved dinitrophenylhydrazine assay. *J. Neurochem.*, **1998**, *70*(1), 268-275. [PMID: 9422371]
- [122] Napolitano, A.; Crescenzi, O.; Pezzella, A.; Prota, G. Generation of the neurotoxin 6-hydroxydopamine by peroxidase/H₂O₂ oxidation of dopamine. *J. Med. Chem.*, **1995**, *38*(6), 917-922. [PMID: 7699708]
- [123] Orth, M.; Tabrizi, S.J. Models of Parkinson's disease. *Mov. Disord.*, **2003**, *18*(7), 729-737. [PMID: 12815651]
- [124] Pezzella, A.; d'Ischia, M.; Napolitano, A.; Misuraca, G.; Prota, G. Iron-mediated generation of the neurotoxin 6-hydroxydopamine quinone by reaction of fatty acid hydroperoxides with dopamine: a possible contributory mechanism for neuronal degeneration in Parkinson's disease. *J. Med. Chem.*, **1997**, *40*(14), 2211-2216. [PMID: 9216840]
- [125] Kaur, D.; Andersen, J. Does cellular iron dysregulation play a causative role in Parkinson's disease? *Ageing Res. Rev.*, **2004**, *3*(3), 327-343. [PMID: 15231240]
- [126] Nunomura, A.; Chiba, S.; Kosaka, K.; Takeda, A.; Castellani, R.J.; Smith, M.A.; Perry, G. Neuronal RNA oxidation is a prominent feature of dementia with Lewy bodies. *Neuroreport*, **2002**, *13*(16), 2035-2039. [PMID: 12438921]
- [127] Salazar, J.; Mena, N.; Núñez, M.T. Iron dyshomeostasis in Parkinson's disease. *J. Neural Transm. Suppl.*, **2006**, (71), 205-213. [PMID: 17447431]
- [128] Sian-Hülsmann, J.; Mandel, S.; Youdim, M.B.; Riederer, P. The relevance of iron in the pathogenesis of Parkinson's disease. *J. Neurochem.*, **2011**, *118*(6), 939-957. [PMID: 21138437]
- [129] Youdim, M.B.; Stephenson, G.; Ben Shachar, D. Ironing iron out in Parkinson's disease and other neurodegenerative diseases with iron chelators: a lesson from 6-hydroxydopamine and iron chelators, desferal and VK-28. *Ann. N. Y. Acad. Sci.*, **2004**, *1012*, 306-325. [PMID: 15105275]
- [130] Taylor, D.M.; Gibbs, B.F.; Kabashi, E.; Minotti, S.; Durham, H.D.; Agar, J.N. Tryptophan 32 potentiates aggregation and cytotoxicity of a copper/zinc superoxide dismutase mutant associated with familial amyotrophic lateral sclerosis. *J. Biol. Chem.*, **2007**, *282*(22), 16329-16335. [PMID: 17389599]
- [131] Ferrante, R.J.; Browne, S.E.; Shinobu, L.A.; Bowling, A.C.; Baik, M.J.; MacGarvey, U.; Kowall, N.W.; Brown, R.H., Jr; Beal, M.F. Evidence of increased oxidative damage in both sporadic and familial amyotrophic lateral sclerosis. *J. Neurochem.*, **1997**, *69*(5), 2064-2074. [PMID: 9349552]

- [132] Kikuchi, S.; Shinpo, K.; Ogata, A.; Tsuji, S.; Takeuchi, M.; Makita, Z.; Tashiro, K. Detection of N epsilon-(carboxymethyl)lysine (CML) and non-CML advanced glycation end-products in the anterior horn of amyotrophic lateral sclerosis spinal cord. *Amyotroph. Lateral Scler. Other Motor Neuron Disord.*, **2002**, *3*(2), 63-68. [PMID: 12215227]
- [133] Pedersen, W.A.; Fu, W.; Keller, J.N.; Markesbery, W.R.; Appel, S.; Smith, R.G.; Kasarskis, E.; Mattson, M.P. Protein modification by the lipid peroxidation product 4-hydroxynonenal in the spinal cords of amyotrophic lateral sclerosis patients. *Ann. Neurol.*, **1998**, *44*(5), 819-824. [PMID: 9818940]
- [134] Calabrese, V.; Raffaele, R.; Cosentino, E.; Rizza, V. Changes in cerebrospinal fluid levels of malondialdehyde and glutathione reductase activity in multiple sclerosis. *Int. J. Clin. Pharmacol. Res.*, **1994**, *14*(4), 119-123. [PMID: 7607784]
- [135] Ferretti, G.; Bacchetti, T.; Principi, F.; Di Ludovico, F.; Viti, B.; Angeleri, V.A.; Danni, M.; Provinciali, L. Increased levels of lipid hydroperoxides in plasma of patients with multiple sclerosis: a relationship with paraoxonase activity. *Mult. Scler.*, **2005**, *11*(6), 677-682. [PMID: 16320727]
- [136] Karg, E.; Klivényi, P.; Németh, I.; Bencsik, K.; Pintér, S.; Vécsei, L. Nonenzymatic antioxidants of blood in multiple sclerosis. *J. Neurol.*, **1999**, *246*(7), 533-539. [PMID: 10463352]
- [137] Smith, K.J.; Kapoor, R.; Felts, P.A. Demyelination: the role of reactive oxygen and nitrogen species. *Brain Pathol.*, **1999**, *9*(1), 69-92. [PMID: 9989453]
- [138] Gilgun-Sherki, Y.; Melamed, E.; Offen, D. The role of oxidative stress in the pathogenesis of multiple sclerosis: the need for effective antioxidant therapy. *J. Neurol.*, **2004**, *251*(3), 261-268. [PMID: 15015004]
- [139] Goebel, H.H.; Heipertz, R.; Scholz, W.; Iqbal, K.; Tellez-Nagel, I. Juvenile Huntington chorea: clinical, ultrastructural, and biochemical studies. *Neurology*, **1978**, *28*(1), 23-31. [PMID: 145549]
- [140] Tellez-Nagel, I.; Johnson, A.B.; Terry, R.D. Studies on brain biopsies of patients with Huntington's chorea. *J. Neuropathol. Exp. Neurol.*, **1974**, *33*(2), 308-332. [PMID: 4150800]
- [141] Browne, S.E.; Bowling, A.C.; MacGarvey, U.; Baik, M.J.; Berger, S.C.; Muqit, M.M.; Bird, E.D.; Beal, M.F. Oxidative damage and metabolic dysfunction in Huntington's disease: selective vulnerability of the basal ganglia. *Ann. Neurol.*, **1997**, *41*(5), 646-653. [PMID: 9153527]
- [142] Dong, H.; Zhang, X.; Qian, Y. Mast cells and neuroinflammation. *Med. Sci. Monit. Basic Res.*, **2014**, *20*, 200-206. [PMID: 25529562]
- [143] Louveau, A.; Smirnov, I.; Keyes, T.J.; Eccles, J.D.; Rouhani, S.J.; Peske, J.D.; Derecki, N.C.; Castle, D.; Mandell, J.W.; Lee, K.S.; Harris, T.H.; Kipnis, J. Structural and functional features of central nervous system lymphatic vessels. *Nature*, **2015**, *523*(7560), 337-341. [PMID: 26030524]
- [144] Gendelman, H.E. Neural immunity: friend or foe? *J. Neurovirol.*, **2002**, *8*(6), 474-479. [PMID: 12476342]
- [145] Bal-Price, A.; Matthias, A.; Brown, G.C. Stimulation of the NADPH oxidase in activated rat microglia removes nitric oxide but induces peroxynitrite production. *J. Neurochem.*, **2002**, *80*(1), 73-80. [PMID: 11796745]
- [146] Mander, P.K.; Jekabsone, A.; Brown, G.C. Microglia proliferation is regulated by hydrogen peroxide from NADPH oxidase. *J. Immunol.*, **2006**, *176*(2), 1046-1052. [PMID: 16393992]
- [147] Jekabsone, A.; Mander, P.K.; Tickler, A.; Sharpe, M.; Brown, G.C. Fibrillar beta-amyloid peptide Aβ1-40 activates microglial proliferation via stimulating TNF-α release and H₂O₂ derived from NADPH oxidase: a cell culture study. *J. Neuroinflammation*, **2006**, *3*, 24. [PMID: 16959029]
- [148] Pawate, S.; Shen, Q.; Fan, F.; Bhat, N.R. Redox regulation of glial inflammatory response to lipopolysaccharide and interferon-γ. *J. Neurosci. Res.*, **2004**, *77*(4), 540-551. [PMID: 15264224]
- [149] Block, M.L.; Zecca, L.; Hong, J.S. Microglia-mediated neurotoxicity: uncovering the molecular mechanisms. *Nat. Rev. Neurosci.*, **2007**, *8*(1), 57-69. [PMID: 17180163]
- [150] Heneka, M.T.; Feinstein, D.L. Expression and function of inducible nitric oxide synthase in neurons. *J. Neuroimmunol.*, **2001**, *114*(1-2), 8-18. [PMID: 11240010]
- [151] Murphy, S. Production of nitric oxide by glial cells: regulation and potential roles in the CNS. *Glia*, **2000**, *29*(1), 1-13. [PMID: 10594918]
- [152] Bal-Price, A.; Brown, G.C. Inflammatory neurodegeneration mediated by nitric oxide from activated glia-inhibiting neuronal respiration, causing glutamate release and excitotoxicity. *J. Neurosci.*, **2001**, *21*(17), 6480-6491. [PMID: 11517237]
- [153] Brown, G.C.; Cooper, C.E. Nanomolar concentrations of nitric oxide reversibly inhibit synaptosomal respiration by competing with oxygen at cytochrome oxidase. *FEBS Lett.*, **1994**, *356*(2-3), 295-298. [PMID: 7805858]
- [154] Mander, P.; Borutaite, V.; Moncada, S.; Brown, G.C. Nitric oxide from inflammatory-activated glia synergizes with hypoxia to induce neuronal death. *J. Neurosci. Res.*, **2005**, *79*(1-2), 208-215. [PMID: 15558752]
- [155] Mander, P.; Brown, G.C. Activation of microglial NADPH oxidase is synergistic with glial iNOS expression in inducing neuronal death: a dual-key mechanism of inflammatory neurodegeneration. *J. Neuroinflammation*, **2005**, *2*, 20. [PMID: 16156895]
- [156] Liu, X.; Miller, M.J.; Joshi, M.S.; Thomas, D.D.; Lancaster, J.R., Jr. Accelerated reaction of nitric oxide with O₂ within the hydrophobic interior of biological membranes. *Proc. Natl. Acad. Sci. USA*, **1998**, *95*(5), 2175-2179. [PMID: 9482858]
- [157] Santolini, J.; Meade, A.L.; Stuehr, D.J. Differences in three kinetic parameters underpin the unique catalytic profiles of nitric-oxide synthases I, II, and III. *J. Biol. Chem.*, **2001**, *276*(52), 48887-48898. [PMID: 11684690]
- [158] Schwab, C.; McGeer, P.L. Inflammatory aspects of Alzheimer disease and other neurodegenerative disorders. *J. Alzheimers Dis.*, **2008**, *13*(4), 359-369. [PMID: 18487845]
- [159] Jacobson, J.; Duchon, M.R. Mitochondrial oxidative stress and cell death in astrocytes—requirement for stored Ca²⁺ and sustained opening of the permeability transition pore. *J. Cell Sci.*, **2002**, *115*(Pt 6), 1175-1188. [PMID: 11884517]
- [160] Murphy, M.P. Nitric oxide and cell death. *Biochim. Biophys. Acta*, **1999**, *1411*(2-3), 401-414. [PMID: 10320672]
- [161] Szabó, C.; Zingarelli, B.; O'Connor, M.; Salzman, A.L. DNA strand breakage, activation of poly (ADP-ribose) synthetase, and cellular energy depletion are involved in the cytotoxicity of macrophages and smooth muscle cells exposed to peroxynitrite. *Proc. Natl. Acad. Sci. USA*, **1996**, *93*(5), 1753-1758. [PMID: 8700830]
- [162] Frieden, M.; James, D.; Castelbou, C.; Danckaert, A.; Martinou, J.C.; Demarex, N. Ca²⁺ homeostasis during mitochondrial fragmentation and perinuclear clustering induced by hFis1. *J. Biol. Chem.*, **2004**, *279*(21), 22704-22714. [PMID: 15024001]
- [163] Szabadkai, G.; Simoni, A.M.; Chami, M.; Wieckowski, M.R.; Youle, R.J.; Rizzuto, R. Drp-1-dependent division of the mitochondrial network blocks intraorganellar Ca²⁺ waves and protects against Ca²⁺-mediated apoptosis. *Mol. Cell*, **2004**, *16*(1), 59-68. [PMID: 15469822]
- [164] Kim, H.R.; Won, S.J.; Fabian, C.; Kang, M.G.; Szardenings, M.; Shin, M.G. Mitochondrial DNA aberrations and pathophysiological implications in hematopoietic diseases, chronic inflammatory diseases, and cancers. *Ann. Lab. Med.*, **2015**, *35*(1), 1-14. [PMID: 25553274]
- [165] Andersen, J.K. Oxidative stress in neurodegeneration: cause or consequence? *Nat. Med.*, **2004**, *10*(Suppl.), S18-S25. [PMID: 15298006]
- [166] Kalra, E.K. Nutraceutical—definition and introduction. *AAPS PharmSci*, **2003**, *5*(3), E25. [PMID: 14621960]
- [167] Brower, V. Nutraceuticals: poised for a healthy slice of the healthcare market? *Nat. Biotechnol.*, **1998**, *16*(8), 728-731. [PMID: 9702769]
- [168] Prasad, C. Improving mental health through nutrition: the future. *Nutr. Neurosci.*, **2001**, *4*(4), 251-272. [PMID: 11842893]
- [169] Hardy, G.; Hardy, I.; Ball, P.A. Nutraceuticals—a pharmaceutical viewpoint: part II. *Curr. Opin. Clin. Nutr. Metab. Care*, **2003**, *6*(6), 661-671. [PMID: 14557798]
- [170] Dadhania, V.P.; Tripathi, D.N.; Vikram, A.; Ramarao, P.; Jena, G.B. Intervention of alpha-lipoic acid ameliorates methotrexate-induced oxidative stress and genotoxicity: A study in rat intestine. *Chem. Biol. Interact.*, **2010**, *183*(1), 85-97. [PMID: 19900424]
- [171] Kumar, A.; Sharma, S.S. NF-κappaB inhibitory action of resveratrol: a probable mechanism of neuroprotection in experimental diabetic neuropathy. *Biochem. Biophys. Res. Commun.*, **2010**, *394*(2), 360-365. [PMID: 20211601]

- [172] Prakash, A.; Kumar, A. Implicating the role of lycopene in restoration of mitochondrial enzymes and BDNF levels in β -amyloid induced Alzheimer's disease. *Eur. J. Pharmacol.*, **2014**, *741*, 104-111. [PMID: 25066110]
- [173] Soto-Urquieta, M.G.; López-Briones, S.; Pérez-Vázquez, V.; Saavedra-Molina, A.; González-Hernández, G.A.; Ramírez-Emiliano, J. Curcumin restores mitochondrial functions and decreases lipid peroxidation in liver and kidneys of diabetic db/db mice. *Biol. Res.*, **2014**, *47*(1), 74. [PMID: 25723052]
- [174] Schroeter, H.; Boyd, C.; Spencer, J.P.; Williams, R.J.; Cadenas, E.; Rice-Evans, C. MAPK signaling in neurodegeneration: influences of flavonoids and of nitric oxide. *Neurobiol. Aging*, **2002**, *23*(5), 861-880. [PMID: 12392791]
- [175] Williams, R.J.; Spencer, J.P.; Rice-Evans, C. Flavonoids: antioxidants or signalling molecules? *Free Radic. Biol. Med.*, **2004**, *36*(7), 838-849. [PMID: 15019969]
- [176] Nicolas, C.S.; Amici, M.; Bortolotto, Z.A.; Doherty, A.; Csaba, Z.; Fafouri, A.; Dournaud, P.; Gressens, P.; Collingridge, G.L.; Peineau, S. The role of JAK-STAT signaling within the CNS. *JAK-STAT*, **2013**, *2*(1), e22925. [PMID: 24058789]
- [177] Kim, H.Y.; Park, E.J.; Joe, E.H.; Jou, I. Curcumin suppresses Janus kinase-STAT inflammatory signaling through activation of Src homology 2 domain-containing tyrosine phosphatase 2 in brain microglia. *J. Immunol.*, **2003**, *171*(11), 6072-6079. [PMID: 14634121]
- [178] Spencer, J.P. Flavonoids: modulators of brain function? *Br. J. Nutr.*, **2008**, *99*(E Suppl 1), ES60-77.
- [179] Peng, P.H.; Chiou, L.F.; Chao, H.M.; Lin, S.; Chen, C.F.; Liu, J.H.; Ko, M.L. Effects of epigallocatechin-3-gallate on rat retinal ganglion cells after optic nerve axotomy. *Exp. Eye Res.*, **2010**, *90*(4), 528-534. [PMID: 20114044]
- [180] Craft, S.; Watson, G.S. Insulin and neurodegenerative disease: shared and specific mechanisms. *Lancet Neurol.*, **2004**, *3*(3), 169-178. [PMID: 14980532]
- [181] Bahadoran, Z.; Mirmiran, P.; Azizi, F. Dietary polyphenols as potential nutraceuticals in management of diabetes: a review. *J. Diabetes Metab. Disord.*, **2013**, *12*(1), 43. [PMID: 23938049]
- [182] Aliev, G.; Palacios, H.H.; Walrafen, B.; Lipsitt, A.E.; Obrenovich, M.E.; Morales, L. Brain mitochondria as a primary target in the development of treatment strategies for Alzheimer disease. *Int. J. Biochem. Cell Biol.*, **2009**, *41*(10), 1989-2004. [PMID: 19703659]
- [183] Boveris, A.; Navarro, A. Brain mitochondrial dysfunction in aging. *IUBMB Life*, **2008**, *60*(5), 308-314. [PMID: 18421773]
- [184] Jalali-Nadoushan, M.; Roghani, M. Alpha-lipoic acid protects against 6-hydroxydopamine-induced neurotoxicity in a rat model of hemi-parkinsonism. *Brain Res.*, **2013**, *1505*, 68-74. [PMID: 23415934]
- [185] Melli, G.; Taiana, M.; Camozzi, F.; Triolo, D.; Podini, P.; Quattrini, A.; Taroni, F.; Lauria, G. Alpha-lipoic acid prevents mitochondrial damage and neurotoxicity in experimental chemotherapy neuropathy. *Exp. Neurol.*, **2008**, *214*(2), 276-284. [PMID: 18809400]
- [186] Müller, U.; Krieglstein, J. Prolonged pretreatment with alpha-lipoic acid protects cultured neurons against hypoxic, glutamate-, or iron-induced injury. *J. Cereb. Blood Flow Metab.*, **1995**, *15*(4), 624-630. [PMID: 7790411]
- [187] Prehn, J.H.; Karkoutly, C.; Nuglisch, J.; Peruche, B.; Krieglstein, J. Dihyrolipoate reduces neuronal injury after cerebral ischemia. *J. Cereb. Blood Flow Metab.*, **1992**, *12*(1), 78-87. [PMID: 1345759]
- [188] Salinthonne, S.; Yadav, V.; Bourdette, D.N.; Carr, D.W. Lipoic acid: a novel therapeutic approach for multiple sclerosis and other chronic inflammatory diseases of the CNS. *Endocr. Metab. Immune Disord. Drug Targets*, **2008**, *8*(2), 132-142. [PMID: 18537699]
- [189] Zhang, L.; Xing, G.Q.; Barker, J.L.; Chang, Y.; Maric, D.; Ma, W.; Li, B.S.; Rubinow, D.R. Alpha-lipoic acid protects rat cortical neurons against cell death induced by amyloid and hydrogen peroxide through the Akt signalling pathway. *Neurosci. Lett.*, **2001**, *312*(3), 125-128. [PMID: 11602326]
- [190] Mijnhout, G.S.; Kollen, B.J.; Alkhalaf, A.; Kleefstra, N.; Bilo, H.J. Alpha lipoic Acid for symptomatic peripheral neuropathy in patients with diabetes: a meta-analysis of randomized controlled trials. *Int. J. Endocrinol.*, **2012**, *2012*, 456279.
- [191] Guo, Y.; Jones, D.; Palmer, J.L.; Forman, A.; Dakhil, S.R.; Velasco, M.R.; Weiss, M.; Gilman, P.; Mills, G.M.; Noga, S.J.; Eng, C.; Overman, M.J.; Fisch, M.J. Oral alpha-lipoic acid to prevent chemotherapy-induced peripheral neuropathy: a randomized, double-blind, placebo-controlled trial. *Support. Care Cancer*, **2014**, *22*(5), 1223-1231. [PMID: 24362907]
- [192] Kim, J.H.; Choi, W.; Lee, J.H.; Jeon, S.J.; Choi, Y.H.; Kim, B.W.; Chang, H.I.; Nam, S.W. Astaxanthin inhibits H₂O₂-mediated apoptotic cell death in mouse neural progenitor cells via modulation of P38 and MEK signaling pathways. *J. Microbiol. Biotechnol.*, **2009**, *19*(11), 1355-1363. [PMID: 19996687]
- [193] Lu, Y.P.; Liu, S.Y.; Sun, H.; Wu, X.M.; Li, J.J.; Zhu, L. Neuroprotective effect of astaxanthin on H₂O₂-induced neurotoxicity *in vitro* and on focal cerebral ischemia *in vivo*. *Brain Res.*, **2010**, *1360*, 40-48. [PMID: 20846510]
- [194] Cleren, C.; Yang, L.; Lorenzo, B.; Calingasan, N.Y.; Schomer, A.; Sireci, A.; Wille, E.J.; Beal, M.F. Therapeutic effects of coenzyme Q10 (CoQ10) and reduced CoQ10 in the MPTP model of Parkinsonism. *J. Neurochem.*, **2008**, *104*(6), 1613-1621. [PMID: 17973981]
- [195] Shults, C.W. Coenzyme Q10 in neurodegenerative diseases. *Curr. Med. Chem.*, **2003**, *10*(19), 1917-1921. [PMID: 12871093]
- [196] Spindler, M.; Beal, M.F.; Henchcliffé, C. Coenzyme Q10 effects in neurodegenerative disease. *Neuropsychiatr. Dis. Treat.*, **2009**, *5*, 597-610. [PMID: 19966907]
- [197] Ferrante, K.L.; Shefner, J.; Zhang, H.; Betensky, R.; O'Brien, M.; Yu, H.; Fantasia, M.; Taft, J.; Beal, M.F.; Traynor, B.; Newhall, K.; Donofrio, P.; Caress, J.; Ashburn, C.; Freiberg, B.; O'Neill, C.; Paladenech, C.; Walker, T.; Pestronk, A.; Abrams, B.; Florence, J.; Renna, R.; Schierbecker, J.; Malkus, B.; Cudkowicz, M. Tolerance of high-dose (3,000 mg/day) coenzyme Q10 in ALS. *Neurology*, **2005**, *65*(11), 1834-1836. [PMID: 16344537]
- [198] Storch, A.; Jost, W.H.; Vieregge, P.; Spiegel, J.; Greulich, W.; Durner, J.; Müller, T.; Kupsch, A.; Henningsen, H.; Oertel, W.H.; Fuchs, G.; Kuhn, W.; Niklowitz, P.; Koch, R.; Herting, B.; Reichmann, H. Randomized, double-blind, placebo-controlled trial on symptomatic effects of coenzyme Q(10) in Parkinson disease. *Arch. Neurol.*, **2007**, *64*(7), 938-944. [PMID: 17502459]
- [199] Dutta, K.; Ghosh, D.; Basu, A. Curcumin protects neuronal cells from Japanese encephalitis virus-mediated cell death and also inhibits infective viral particle formation by dysregulation of ubiquitin-proteasome system. *J. Neuroimmune Pharmacol.*, **2009**, *4*(3), 328-337. [PMID: 19434500]
- [200] Giri, R.K.; Rajagopal, V.; Kalra, V.K. Curcumin, the active constituent of turmeric, inhibits amyloid peptide-induced cytokine gene expression and CCR5-mediated chemotaxis of THP-1 monocytes by modulating early growth response-1 transcription factor. *J. Neurochem.*, **2004**, *91*(5), 1199-1210. [PMID: 15569263]
- [201] Kelsey, N.A.; Wilkins, H.M.; Linseman, D.A. Nutraceutical antioxidants as novel neuroprotective agents. *Molecules*, **2010**, *15*(11), 7792-7814. [PMID: 21060289]
- [202] Schmitz, K.; Barthelmes, J.; Stolz, L.; Beyer, S.; Diehl, O.; Tegeder, I. "Disease modifying nutraceuticals" for multiple sclerosis. *Pharmacol. Ther.*, **2015**, *148*, 85-113. [PMID: 25435020]
- [203] Yu, S.; Zheng, W.; Xin, N.; Chi, Z.H.; Wang, N.Q.; Nie, Y.X.; Feng, W.Y.; Wang, Z.Y. Curcumin prevents dopaminergic neuronal death through inhibition of the c-Jun N-terminal kinase pathway. *Rejuvenation Res.*, **2010**, *13*(1), 55-64. [PMID: 20230279]
- [204] Zhao, J.; Yu, S.; Zheng, W.; Feng, G.; Luo, G.; Wang, L.; Zhao, Y. Curcumin improves outcomes and attenuates focal cerebral ischemic injury via antiapoptotic mechanisms in rats. *Neurochem. Res.*, **2010**, *35*(3), 374-379. [PMID: 19774461]
- [205] Kraft, A.D.; Johnson, D.A.; Johnson, J.A. Nuclear factor E2-related factor 2-dependent antioxidant response element activation by tert-butylhydroquinone and sulforaphane occurring preferentially in astrocytes conditions neurons against oxidative insult. *J. Neurosci.*, **2004**, *24*(5), 1101-1112. [PMID: 14762128]
- [206] Brewer, G.J.; Torricelli, J.R.; Lindsey, A.L.; Kunz, E.Z.; Neuman, A.; Fisher, D.R.; Joseph, J.A. Age-related toxicity of amyloid-beta associated with increased pERK and pCREB in primary hippocampal neurons: reversal by blueberry extract. *J. Nutr. Biochem.*, **2010**, *21*(10), 991-998. [PMID: 19954954]
- [207] Fuentealba, J.; Dibarrat, A.J.; Fuentes-Fuentes, M.C.; Saez-Orellana, F.; Quiñones, K.; Guzmán, L.; Perez, C.; Becerra, J.; Aguayo, L.G. Synaptic failure and adenosine triphosphate

- imbalance induced by amyloid- β aggregates are prevented by blueberry-enriched polyphenols extract. *J. Neurosci. Res.*, **2011**, *89*(9), 1499-1508. [PMID: 21647937]
- [208] Petrozzi, L.; Ricci, G.; Giglioli, N.J.; Siciliano, G.; Mancuso, M. Mitochondria and neurodegeneration. *Biosci. Rep.*, **2007**, *27*(1-3), 87-104. [PMID: 17486441]
- [209] Shukitt-Hale, B.; Lau, F.C.; Carey, A.N.; Galli, R.L.; Spangler, E.L.; Ingram, D.K.; Joseph, J.A. Blueberry polyphenols attenuate kainic acid-induced decrements in cognition and alter inflammatory gene expression in rat hippocampus. *Nutr. Neurosci.*, **2008**, *11*(4), 172-182. [PMID: 18681986]
- [210] Ogle, W.O.; Speisman, R.B.; Ormerod, B.K. Potential of treating age-related depression and cognitive decline with nutraceutical approaches: a mini-review. *Gerontology*, **2013**, *59*(1), 23-31. [PMID: 22947921]
- [211] Pocerich, C.B.; Lange, M.L.; Sultana, R.; Butterfield, D.A. Nutritional approaches to modulate oxidative stress in Alzheimer's disease. *Curr. Alzheimer Res.*, **2011**, *8*(5), 452-469. [PMID: 21605052]
- [212] Lee, H.J.; Cho, H.S.; Park, E.; Kim, S.; Lee, S.Y.; Kim, C.S.; Kim, D.K.; Kim, S.J.; Chun, H.S. Rosmarinic acid protects human dopaminergic neuronal cells against hydrogen peroxide-induced apoptosis. *Toxicology*, **2008**, *250*(2-3), 109-115. [PMID: 18644421]
- [213] Qiao, S.; Li, W.; Tsubouchi, R.; Haneda, M.; Murakami, K.; Takeuchi, F.; Nisimoto, Y.; Yoshino, M. Rosmarinic acid inhibits the formation of reactive oxygen and nitrogen species in RAW264.7 macrophages. *Free Radic. Res.*, **2005**, *39*(9), 995-1003. [PMID: 16087481]
- [214] Peng, Q.; Buz'Zard, A.R.; Lau, B.H. Neuroprotective effect of garlic compounds in amyloid-beta peptide-induced apoptosis *in vitro*. *Med. Sci. Monit.*, **2002**, *8*(8), BR328-BR337. [PMID: 12165737]
- [215] Kabuto, H.; Tada, M.; Kohno, M. Eugenol [2-methoxy-4-(2-propenyl)phenol] prevents 6-hydroxydopamine-induced dopamine depression and lipid peroxidation inductivity in mouse striatum. *Biol. Pharm. Bull.*, **2007**, *30*(3), 423-427. [PMID: 17329831]
- [216] Frisardi, V.; Panza, F.; Solfrizzi, V.; Seripa, D.; Pilotto, A. Plasma lipid disturbances and cognitive decline. *J. Am. Geriatr. Soc.*, **2010**, *58*(12), 2429-2430. [PMID: 21143446]
- [217] Sun, G.Y.; Xu, J.; Jensen, M.D.; Simonyi, A. Phospholipase A2 in the central nervous system: implications for neurodegenerative diseases. *J. Lipid Res.*, **2004**, *45*(2), 205-213. [PMID: 14657205]
- [218] Fragoso, Y.D.; Stoney, P.N.; McCaffery, P.J. The evidence for a beneficial role of vitamin A in multiple sclerosis. *CNS Drugs*, **2014**, *28*(4), 291-299. [PMID: 24557746]
- [219] Gianforcaro, A.; Hamadeh, M.J. Vitamin D as a potential therapy in amyotrophic lateral sclerosis. *CNS Neurosci. Ther.*, **2014**, *20*(2), 101-111. [PMID: 24428861]
- [220] Betti, M.; Minelli, A.; Ambrogini, P.; Ciuffoli, S.; Viola, V.; Galli, F.; Canonico, B.; Lattanzi, D.; Colombo, E.; Sestili, P.; Cuppini, R. Dietary supplementation with α -tocopherol reduces neuro-inflammation and neuronal degeneration in the rat brain after kainic acid-induced status epilepticus. *Free Radic. Res.*, **2011**, *45*(10), 1136-1142. [PMID: 21749318]
- [221] Pu, H.; Guo, Y.; Zhang, W.; Huang, L.; Wang, G.; Liou, A.K.; Zhang, J.; Zhang, P.; Leak, R.K.; Wang, Y.; Chen, J.; Gao, Y. Omega-3 polyunsaturated fatty acid supplementation improves neurologic recovery and attenuates white matter injury after experimental traumatic brain injury. *J. Cereb. Blood Flow Metab.*, **2013**, *33*(9), 1474-1484. [PMID: 23801244]
- [222] Losi, G.; Puia, G.; Garzon, G.; de Vuono, M.C.; Baraldi, M. Apigenin modulates GABAergic and glutamatergic transmission in cultured cortical neurons. *Eur. J. Pharmacol.*, **2004**, *502*(1-2), 41-46. [PMID: 15464088]
- [223] Zhao, L.; Wang, J.L.; Wang, Y.R.; Fa, X.Z. Apigenin attenuates copper-mediated β -amyloid neurotoxicity through antioxidant, mitochondrion protection and MAPK signal inactivation in an AD cell model. *Brain Res.*, **2013**, *1492*, 33-45. [PMID: 23178511]
- [224] Lee, Y.B.; Lee, H.J.; Won, M.H.; Hwang, I.K.; Kang, T.C.; Lee, J.Y.; Nam, S.Y.; Kim, K.S.; Kim, E.; Cheon, S.H.; Sohn, H.S. Soy isoflavones improve spatial delayed matching-to-place performance and reduce cholinergic neuron loss in elderly male rats. *J. Nutr.*, **2004**, *134*(7), 1827-1831. [PMID: 15226476]
- [225] Lee, H.; Kim, H.J.; Kim, J.M.; Chang, N. Effects of dietary folic acid supplementation on cerebrovascular endothelial dysfunction in rats with induced hyperhomocysteinemia. *Brain Res.*, **2004**, *996*(2), 139-147. [PMID: 14697491]
- [226] Mizuno, M.; Kawamura, H.; Takei, N.; Nawa, H. The anthraquinone derivative Emodin ameliorates neurobehavioral deficits of a rodent model for schizophrenia. *J. Neural Transm (Vienna)*, **2008**, *115*(3), 521-530. [PMID: 18301953]
- [227] Duncan, R.S.; Goad, D.L.; Grillo, M.A.; Kaja, S.; Payne, A.J.; Koulen, P. Control of intracellular calcium signaling as a neuro-protective strategy. *Molecules*, **2010**, *15*(3), 1168-1195. [PMID: 20335972]
- [228] Calkins, M.J.; Johnson, D.A.; Townsend, J.A.; Vargas, M.R.; Dowell, J.A.; Williamson, T.P.; Kraft, A.D.; Lee, J.M.; Li, J.; Johnson, J.A. The Nrf2/ARE pathway as a potential therapeutic target in neurodegenerative disease. *Antioxid. Redox Signal.*, **2009**, *11*(3), 497-508. [PMID: 18717629]
- [229] Han, J.M.; Lee, Y.J.; Lee, S.Y.; Kim, E.M.; Moon, Y.; Kim, H.W.; Hwang, O. Protective effect of sulforaphane against dopaminergic cell death. *J. Pharmacol. Exp. Ther.*, **2007**, *321*(1), 249-256. [PMID: 17259450]
- [230] Seidl, S.E.; Santiago, J.A.; Bilyk, H.; Potashkin, J.A. The emerging role of nutrition in Parkinson's disease. *Front. Aging Neurosci.*, **2014**, *6*, 36. [PMID: 24639650]